



Effects of Chromium (VI) on Haematological Parameters in *Channa punctatus* (Bloch. 1793)

Jyotsna Dixit ^a and Vivek Kumar ^{a*}

^a Department of Zoology, Isabella Thoburn College, Lucknow- 226007, India.

Authors' contributions

This work was carried out in collaboration between both authors. Both authors read and approved the final manuscript.

Article Information

DOI: 10.9734/IJECC/2023/v13i92407

Open Peer Review History:

This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here: <https://www.sdiarticle5.com/review-history/101744>

Original Research Article

Received: 03/05/2023

Accepted: 05/07/2023

Published: 22/07/2023

ABSTRACT

Numerous enterprises discharge their untreated waste water straight into aquatic habitats. In addition to causing water pollution, the poisonous heavy metals like chromium found in that waste water also have an impact on aquatic life. Therefore, the goal of this research was to assess the toxicological effects of chromium (VI) on *Channa punctatus*, a freshwater fish. Chromium is an important metal and also a severe threat to aquatic life. Fish, the dominant creature in the aquatic environment, end up being the main target of these contaminants. Chromium enters fish's bodies via their gills, skin, and digestive tracts as a result of contaminated food. They are eventually carried by the circulation to the organs and tissues, where they accumulate after absorption. Through the food chain, heavy metals ultimately find their way into people. Approximately 120 fishes were subjected to three different amounts of chromium (LC₅₀/5: 15.37 mg/L, LC₅₀/10: 7.68 mg/L, and LC₅₀/20: 3.84 mg/L) in this experiment, as well as a toxicant-free control. The relevant blood samples were taken from the caudal peduncle at intervals of 7, 14, 21, and 28 days in order to determine haematological parameters (haemoglobin, haematocrit, red blood cell count, etc.). As a consequence, several alterations in these parameters were noted, and eventually, fish began to lose their resistance to disease, become anaemic, and weaken their immune systems. While it was

*Corresponding author: E-mail: vivekkumarsb@gmail.com;

stated that throughout exposure periods of 7, 14, 21, and 28 days, Groups 2, 3, and 4 had substantially higher MCH and MCV values than Control (Group 1), it was also observed that Groups 2, 3, and 4 had lower Hb, RBC and PCV values. The primary objective of this research was to increase public awareness of the need to protect aquatic environments, particularly from heavy metal-contamination.

Keywords: Haematological parameters; haemoglobin; *Channa punctatus*; chromium trioxide.

1. INTRODUCTION

Because heavy metals are persistent and non-biodegradable, and because they tend to accumulate in living things, environmentalists are now highly worried about water pollution brought on by heavy metal contamination [1]. Heavy metals in the atmosphere are irreversible, so they either accumulate in biota or leach into groundwater [2]. Heavy metals are essential to many biological processes in aquatic species and are still present in the body in trace amounts, or in amounts that do not surpass 1 µg/g. However, even a little concentration rise causes substantial levels of toxicity in several organs. One of the most typical pollutants in surface and ground water is chromium (Cr). In an aquatic ecosystem, Cr travels through effluents that are primarily released from different sources, including metal finishing, dyeing, and printing industries, leather tanneries, mining, photographic, pharmaceutical, electroplating, ceramic, and textile industries, etc. [3-5]. On the other hand, insufficient treatment of these effluents might lead to Cr (VI) contamination of neighbouring water bodies, where it is often found at concentrations that could be harmful to fish [6,7]. Cr (VI) interacts negatively with the environment due to its capacity to maintain persistence and produce a broad variety of unfavourable effects on biological and ecological systems, including fish and aquatic life [8]. Consuming chromium-tainted aquatic food has raised the danger of human health issues worldwide, but especially in developing nations like India. Fish are thus regarded as one of the most crucial indicators in aquatic systems for tracking trace metal contamination and its potential effects on human health [1]. Since fish make up the majority of the aquatic creatures in this system, they are also the most susceptible to these poisons. Chromium is ingested by fish either via their gills or their stomachs, where it circulates throughout the body before being subsequently deposited in different organs [9]. Heavy metal contamination reportedly reduced fish production [10].

In many fish species, haematology serves as a gauge of overall health. Fish blood's haematological indicators may tell us a lot about how physiologically a fish reacts to changes in its environment. For many years, metal pollution in the aquatic environment has been monitored using fish haematological indicators [11]. Blood indices are thus considered pathophysiological markers of the whole organism in order to assess whether fish have suffered structural and functional damage as a consequence of toxic exposure [12]. Fish exposed to heavy metals may have changes in their blood parameters. Diverse impacts on the levels of Hb and Hct have diverse effects on the blood's capacity to transport oxygen depending on the kind of heavy metal, the species, and the time of exposure. Numerous haematological indicators, including haemoglobin (Hb), haemoglobin concentration (Hct), red blood cells (RBCs), and others, have been employed in aquatic settings as indicators of metal pollution and are routinely used to assess the functional status of the bloodstream's capacity to supply oxygen [12]. The author chose to focus on chromium among all the heavy metals since there are several tanneries close to the Lucknow region that ultimately dump chromium. Investigating the effects of chromium toxicity on several *Channa punctatus*'s haematological parameters is the main goal of the current investigation. Since environmental toxicants primarily affect fish among aquatic organisms and, as a result of these pollutants, fish health deteriorates over time, an attempt has been made to correlate the impact of these pollutants on the fish's initial health markers, namely, selected haematological parameters, in the current experiment.

2. MATERIALS AND METHODS

2.1 Test Animal

Channa punctatus, a freshwater edible fish, were collected from local lentic habitat of Lucknow with the help of a local fisherman. The fish were brought to the Lucknow University Laboratory of Zoology Department in a plastic container with

water inside that was covered with a cotton rag to keep the fish from escaping. Fish were immersed in a 0.1% KMnO₄ solution for three to five minutes to remove any contaminants from the dermal region. The fish were accustomed to laboratory conditions for 15 days in aquariums with 100 L of fresh tap water prior to the experiment. During the acclimatisation period, fish were fed optimal fish food twice daily. Every aquarium was regularly examined for fish mortality, and those that were found dead were removed from the tanks.

2.2 Test Chemical

The test chemical was made by Merck Specialities Private Limited, Shiv Sagar Estate A (Dr. Annie Besant Road, Worli, Mumbai, 400018), and was purified CrO₃ Chromium (VI) Oxide. All compounds that were employed in different formulations were of the highest commercial grade.

2.3 Experimental Approach

There were two sets used to conduct the experiment. The EPA Probit Analysis Programme was used to calculate the LC₅₀ values for CrO₃ during the course of the first set of experiments, which lasted 96 hours. The fish were split into four groups of 30 fish each for the second set. While fish from groups 1 retained as the control group with no treatment, fish from groups 2, 3, and 4 were given sub-lethal doses of chromium trioxide for 28 days at LC₅₀/20: 3.84 mg/L, LC₅₀/10: 7.68 mg/L, and LC₅₀/5: 15.37 mg/L, respectively. The experimental setup received appropriate aeration. After the study started, mortality was tracked every day, and dead fish were immediately removed.

2.4 Haematological Analysis

After 7, 14, 21, and 28 days respectively, blood samples from the fish that had survived in each tank were obtained using the caudal vein technique for haematological study. The Neubauer haemocytometer was used to count the RBCs and WBCs under a light microscope. Haemoglobin (Hb%) was determined using haemoglobin strips and a digital EasyMate® GHb (Model: ET 232, Hb/Glu double monitoring device, Bioptic Technology Inc., Taiwan 35057). According to Jain [13], the packed cell volume (PCV, %), mean corpuscular volume (MCV, m³), and mean corpuscular

haemoglobin (MCH, pg) were all calculated using the formulae below.

$$\text{PCV (\%)} = \% \text{ Hb} \times 3$$

$$\text{MCV (\mu m}^3\text{)} = (\text{PCV/RBC in millions}) \times 10$$

$$\text{MCH (pg)} = (\% \text{ Hb/RBC in millions}) \times 10$$

2.5 Evaluation of the Data and Statistical Analysis

The standard deviation (SD) was used to represent values. By using probit analysis, the LC₅₀ 96 hrs value of chromium was calculated. A one-way ANOVA was used to analyse the data, and then a post hoc test was used to determine if there were any statistically significant differences between treatment and control values. At a significance level of $p < 0.05$, statistical analysis was carried out using SPSS version 27.0 for Windows (SPSS Inc., Chicago, IL, USA).

3. RESULTS AND DISCUSSION

The freshwater fish *Channa punctatus*' haematological parameters for 7, 14, 21, and 28 days were studied in the present research to determine the impact of sub-lethal amounts of chromium trioxide on them. RBC count, Hb%, PCV, MCH, and MCV were among the variable haematological indicators. After being exposed for 28 days to various concentrations of chromium trioxide, the fish in group 1 (the control) had haemoglobin percentages that ranged from 11.00 ± 0.50 % to 11.20 ± 0.20 %; however, these values dropped to 10.20 ± 0.20 % to 10.80 ± 0.50 % in group 2, 9.60 ± 0.20 % to 10.30 ± 0.50 % in group 3, and 8.20 ± 0.20 % to 9.50 ± 0.30 % in group 4, which indicated a significant difference ($P < 0.05$). Fish in group 4 had the greatest Hb% decline. When compared to control, group 4's RBC count value fell the most. The RBC count ranged from 2.21 ± 0.21 to 2.23 ± 0.36 in group 1, whereas it ranged from 1.00 ± 0.32 to 1.09 ± 0.32 in group 4, which differed significantly ($P < 0.05$). In comparison to group 1, groups 2 and 3 similarly showed a significant difference ($P < 0.05$) in RBC count. The packed cell volume (PCV)% values for groups 2, 3, and 4 were significantly ($P < 0.05$) lower than those for group 1, with group 1 having the highest values, which ranged from 30 ± 1.00 to 30.33 ± 1.60 , and group 4 having the lowest values, which ranged from 25.00 ± 0.50 to 25.50 ± 1.50 . On the other hand, after a 28-day exposure to chromium trioxide, mean

corpuscular haemoglobin (MCH) and mean corpuscular volume (MCV) levels were significantly ($P < 0.05$) higher than controls. MCH values for groups 2, 3, and 4 ranged from 50.45 ± 4.15 to 54.33 ± 4.15 , 65 ± 3.26 to 70.15 ± 1.20 and 70.23 ± 3.88 to 77.55 ± 3.44 , respectively. Group 1 had the lowest MCH

values, which ranged from 33.48 ± 4.68 to 35.48 ± 4.50 . MCV values in group 4 ranged from 202.33 ± 5.78 to 239.45 ± 3.57 , whereas those in group 1 ranged from 82.14 ± 4.26 to 85.23 ± 4.12 . Group 4 had the highest MCV values. Values for groups 2 and 3 were likewise raised in comparison to group 1.

Table 1. Showing variations in haematological parameters of *Channa punctatus* exposed to CrO_3 for different durations

Parameters	Exposure time (days)	Group1 (control) Mean \pm SEM	Group 2 (LC ₅₀ /20) Mean \pm SEM	Group 3 (LC ₅₀ /10) Mean \pm SEM	Group 4 (LC ₅₀ /5) Mean \pm SEM
RBC Count (millions/cumm)	7	2.21 \pm 0.21	2.15 \pm 0.48	1.65 \pm 0.43	1.09 \pm 0.32
	14	2.21 \pm 0.52	2.15 \pm 0.55	1.58 \pm 0.37	1.06 \pm 0.32
	21	2.23 \pm 0.36	2.04 \pm 0.33	1.40 \pm 0.33	1.02 \pm 0.43
	28	2.22 \pm 0.34	2.00 \pm 0.26	1.40 \pm 0.32	1.00 \pm 0.32
Haemoglobin (Hb) (%)	7	11.20 \pm 0.20	10.80 \pm 0.50	10.30 \pm 0.50	9.50 \pm 0.30
	14	11.13 \pm 0.30	10.60 \pm 0.60	10.30 \pm 1.20	9.20 \pm 0.30
	21	11.10 \pm 0.50	10.50 \pm 0.30	10.10 \pm 0.50	8.60 \pm 0.60
	28	11.00 \pm 0.50	10.20 \pm 0.20	9.60 \pm 0.20	8.20 \pm 0.20
Hematocrit (PCV %)	7	30.33 \pm 2.50	29.83 \pm 1.50	27.50 \pm 1.15	25.50 \pm 1.50
	14	30.33 \pm 1.60	29.50 \pm 1.30	27.30 \pm 1.20	25.50 \pm 1.20
	21	30.15 \pm 1.50	29.20 \pm 1.00	27.10 \pm 1.30	25.20 \pm 1.15
	28	30.15 \pm 1.00	29.45 \pm 0.50	26.50 \pm 1.15	25.00 \pm 0.50
MCH (pg)	7	35.48 \pm 4.50	50.55 \pm 4.21	65 \pm 3.26	70.23 \pm 3.88
	14	34.33 \pm 4.25	50.45 \pm 4.15	65.44 \pm 3.50	72.46 \pm 3.40
	21	34.30 \pm 4.15	52.35 \pm 4.30	67.23 \pm 2.77	76.54 \pm 2.67
	28	33.48 \pm 4.68	54.33 \pm 4.15	70.15 \pm 1.20	77.55 \pm 3.44
MCV	7	82.33 \pm 4.46	103.96 \pm 0.97	183.64 \pm 2.34	205.65 \pm 5.66
	14	82.14 \pm 4.26	105.4 \pm 0.92	187.56 \pm 2.66	202.33 \pm 5.78
	21	83.11 \pm 4.33	105.86 \pm 0.96	190.33 \pm 2.87	210.56 \pm 4.78
	28	85.23 \pm 4.12	108.66 \pm 0.99	195.46 \pm 3.48	239.45 \pm 3.57

The data are represented in mean \pm SEM

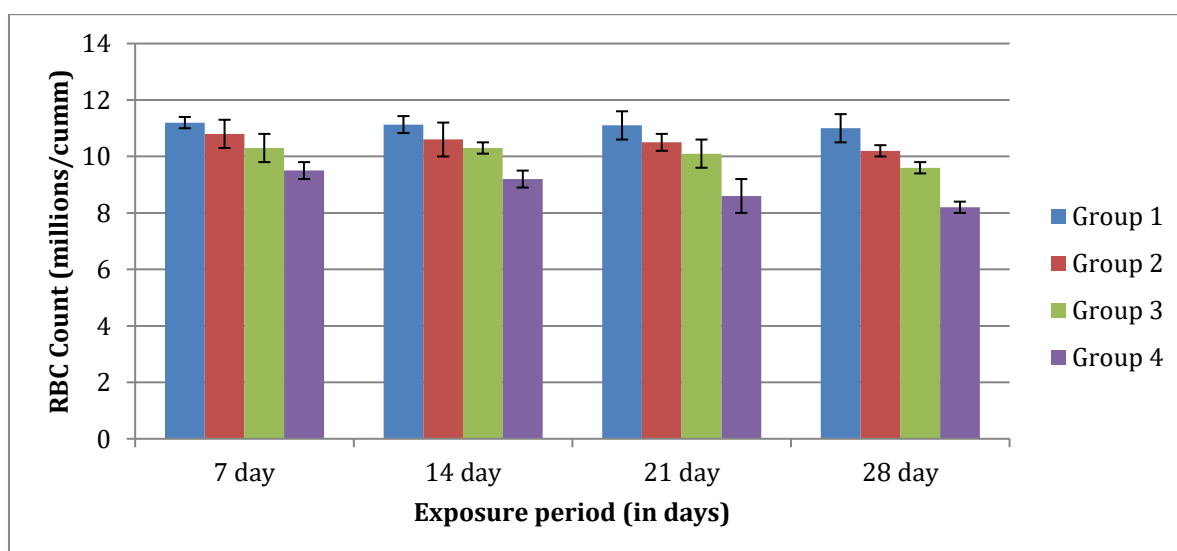


Fig. 1. RBC count values of *C. punctatus* exposed to varying amount of CrO_3

The values are represented in mean \pm SEM

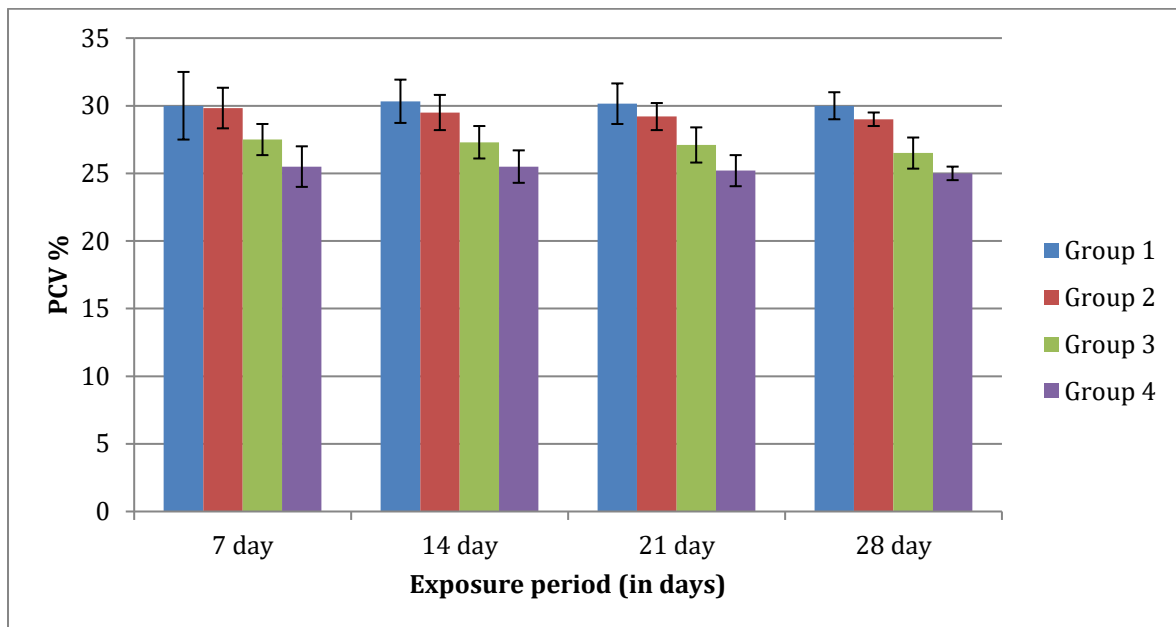


Fig. 2. PCV values of *C. punctatus* exposed to varying amount of CrO₃
The values are represented in mean \pm SEM

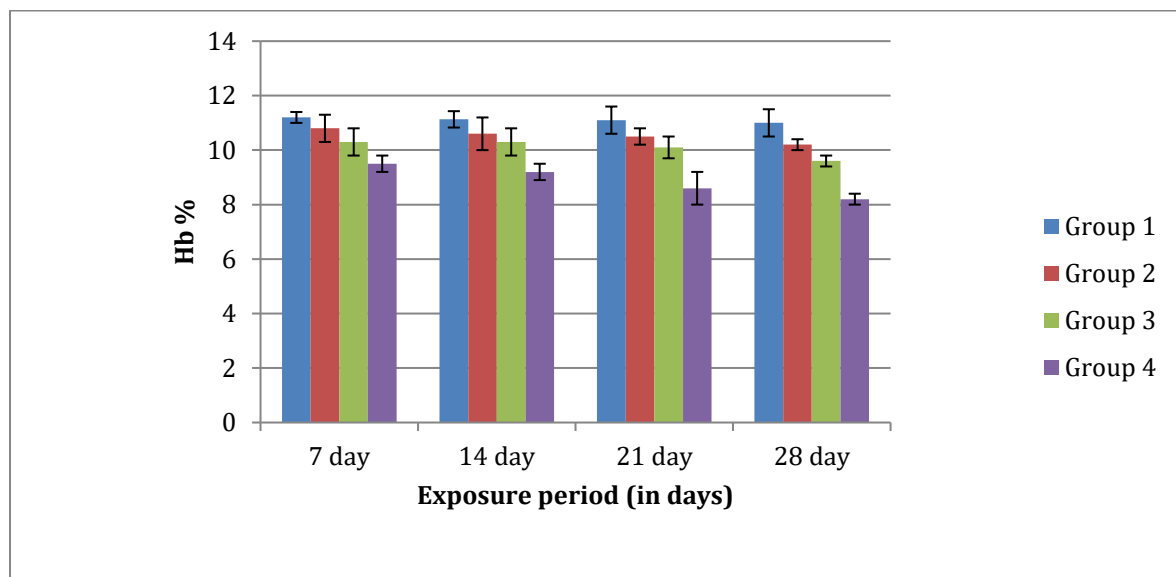


Fig. 3. Hb % values of *C. punctatus* exposed to varying amount of CrO₃
The values are represented in mean \pm SEM

Haematology is used as an indicator of fish health conditions in a range of fish species in order to detect different stress situations such as illness, hypoxia, exposure to metals and pollutants, etc. [14]. The results of this research show that fish erythrocytic parameters are affected by hexavalent chromium. According to Javed et al. [1], the decrease in haematological values might potentially be attributed to heavy metals (pollutants) altering

the structure of RBCs, which prevented more oxygen molecules from binding to Hb, decreased RBC oxygen carrying capacity, decreased Hb levels, and even caused anaemia. According to past studies [15,16], heavy metals may damage erythrocytes, which would explain the significant drop in haemoglobin content. Because of the low haemoglobin level, the fish's vital organs are not getting enough oxygen. Degradation of the erythrocytes and a persistent drop in

haemoglobin concentration in fish exposed to heavy metals may be regarded as pathological conditions that limit oxygen delivery [17]. To further pinpoint the anaemic status of these experimental fish groups, haematocrit measurement was employed. Both haemoglobin levels and Hct% significantly decreased after exposure to the heavy metal chromium, which clearly suggests a haemodilution mechanism that may be brought on by gill injury or poor

osmoregulation. In *Labeo rohita* exposed to chromium, the mean MCV and MCH levels significantly altered as well [18]. A similar discovery in *C. carpio* after exposure to cadmium was reported [19]. The fact that the values for Hb% and haematocrit in *Channa punctatus* exposed to both copper and chromium declined drastically implies that metal causes acute anaemia under risky circumstances, according to Singh [20].

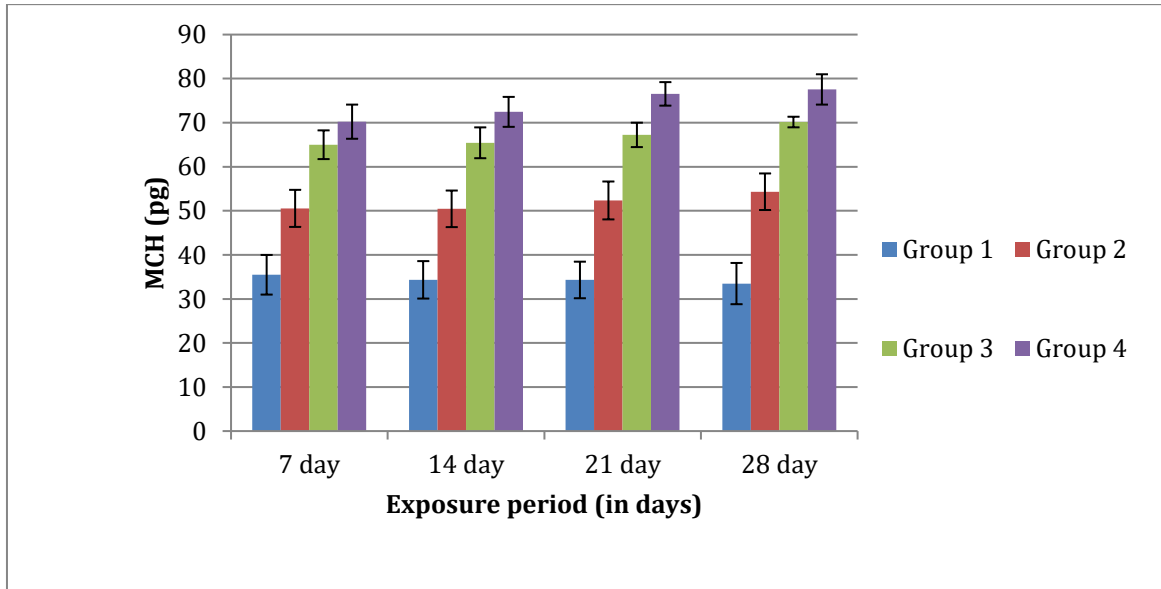


Fig. 4. MCH values of *C. punctatus* exposed varying amount of CrO_3
The values are represented in mean \pm SEM

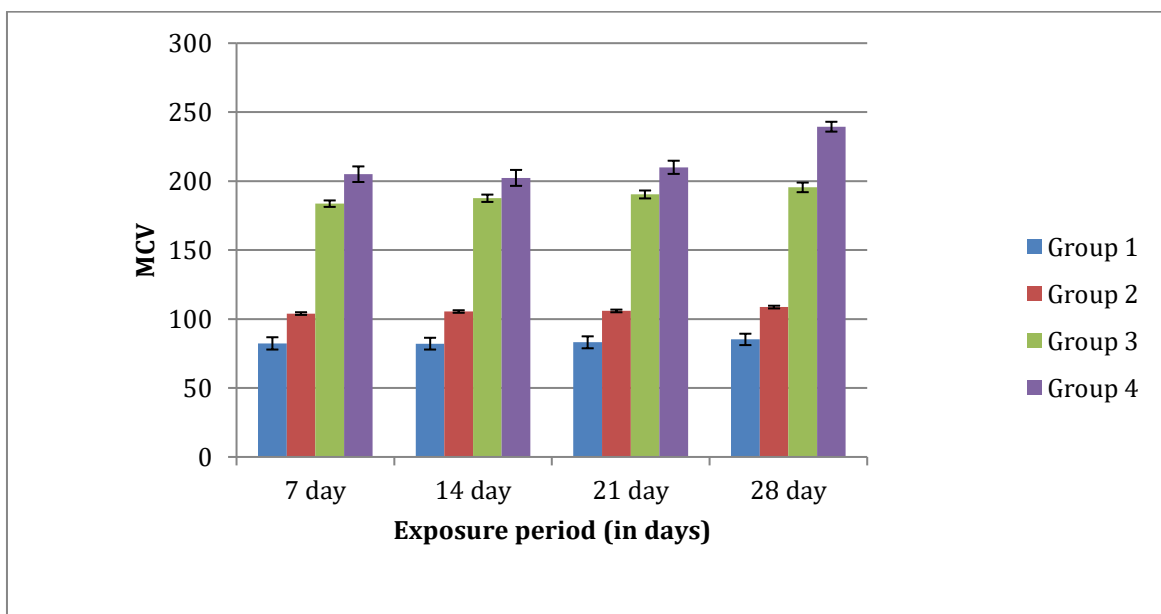


Fig. 5. MCV values of *C. punctatus* exposed varying amount of CrO_3
The values are represented in mean \pm SEM

4. CONCLUSION

Pollutants pose a major danger to aquatic ecosystems. This research shows a clear correlation between Cr (VI) content and fish health. Freshwater fish species are more vulnerable to Cr (VI) in comparison to other aquatic organisms. Haematological measures can be used to measure chromium since they may be helpful in the early diagnosis of fish health conditions before they worsen. It is hoped that, the research community will be benefit from the information on the ecotoxicology and risk assessment of hazardous chromium by the scientific evidence offered in this study, which provides a basis for comprehending the possible influence of chromium on a fish species' haemoglobin. Furthermore, it has been shown that fish exhibit a broad spectrum of chromium-related impacts on their histology, physiology, biochemistry, enzymology, haematology, and genetic make-up. The same negative effects that chromium has on indicator organisms (fish) will ultimately reach humans when it enters the food chain via water and food. Thus, one of the main approaches to solving the issue is to prevent chromium poisoning of aquatic habitats from happening again.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Javed M, Ahmad I, Ahmad A, Usmani N, Ahmad M. Studies on the alterations in Hematological indices, micronuclei induction and pathological marker enzyme activities in *Channa punctatus* (spotted snakehead) perciformes, channidae exposed to thermal power plant effluent. Springer Plus, 2016;5:1–9.
2. Islam MS, Proshad R, Ahmed S. Ecological risk of heavy metals in sediment of an urban river in Bangladesh. Human and Ecological Risk Assessment: An International Journal. 2018;24(3):699–720.
3. GracePavithra K, Jaikumar V, Kumar PS, SundarRajan P. A review on cleaner strategies for chromium industrial wastewater: present research and future perspective. Journal of Cleaner Production. 2019;228:580–593.
4. Liknaw G, Tekalign T, Guya K. Impacts of tannery effluent on environments and human health: A review article. 2017;54. Available:www.iiste.org
5. Ukhurebor KE, Aigbe, UO, Onyancha RB, Nwankwo W, Osibote OA, Paumo HK, Ama OM, Adetunji CO, Siloko IU. Effect of hexavalent chromium on the environment and removal techniques: A review. Journal of Environmental Management. 2021;280: 111809. Available:https://doi.org/10.1016/j.jenvman.2020.111809
6. Bhatia SC. Pollution control in textile industry. CRC Press2017.
7. Lellis B, Fávoro-Polonio CZ, Pamphile JA, Polonio JC. Effects of textile dyes on health and the environment and bioremediation potential of living organisms. Biotechnology Research and Innovation. 2019;3(2):275–290.
8. Ali H, Khan E, Ilahi I. Environmental chemistry and ecotoxicology of hazardous heavy metals: Environmental persistence, toxicity, and bioaccumulation. In Journal of Chemistry. Hindawi Limited. 2019:2019. Available:https://doi.org/10.1155/2019/6730305
9. AL-Taee SK, Al-Mallah KH, Khlsmail H. Review on some heavy metals toxicity on freshwater fishes. Journal of Applied Veterinary Sciences. 2020;5(3):78–86. Available:https://doi.org/10.21608/JAVS
10. Lu Y, Wang R, Zhang Y, Su H, Wang P, Jenkins A, Ferrier RC, Bailey M, Squire G. Ecosystem health towards sustainability. Ecosystem Health and Sustainability. 2015;1(1):1–15. Available:https://doi.org/10.1890/EHS14-0013.1
11. Authman MM. Use of fish as bio-indicator of the effects of heavy metals pollution. Journal of Aquaculture Research & Development. 2015;06(04). Available:https://doi.org/10.4172/2155-9546.1000328
12. Fazio F. Fish hematology analysis as an important tool of aquaculture: A review. In Aquaculture Elsevier B.V. 2019;500:237–242. Available:https://doi.org/10.1016/j.aquaculture.2018.10.030
13. Jain NC. Essentials of veterinary hematology; 1993.
14. Blaxhall PC, Daisley KW. Routine hematological methods for use with fish

- blood. Journal of Fish Biology. 1973;5(6): 771–781.
15. Karuppasamy R. Impact of Phenyl Mercuric Acetate (PMA) on the biomodal respiration in an air breathing fish, *Channa punctatus* (bloch). Journal of Environment and Pollution. 2000;7(4):287–293.
 16. Zutshi B, et al. Alteration in hematology of *Labeo rohita* under stress of pollution from lakes of Bangalore, Karnataka, India. Environmental Monitoring and Assessment. 2010;168:11–19.
 17. Pamila D, et al. Toxic effects of chromium and cobalt on *Sarotherodon mossambicus* (peters). Indian Journal of Environmental Health. 1991;33(2):218–224.
 18. Venkatachalam T, Natarajan AV. Hematological investigation on freshwater teleost *Labeo rohita* (ham.) following aquatic toxicities of Cr (III) and Cr (VI). International Journal of Research in Biosciences. 2014;3(3):1–14.
 19. Koyama J, Ozaki H. Hematological changes of fish exposed to low concentrations of cadmium in the water. Bulletin of the Japanese Society of Scientific Fisheries; 1984.
 20. Singh M. Hematological responses in a freshwater teleost *Channa punctatus* to experimental copper and chromium poisoning. Journal of Environmental Biology. 1995;16(4):339–341.

© 2023 Dixit and Kumar; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:

The peer review history for this paper can be accessed here:
<https://www.sdiarticle5.com/review-history/101744>