



Survey of the Iron Status of Patients with Type 2 Diabetes Mellitus Attending Hospitals in Jos

P. O. Okonkwo^{1,2*} and Z. S. C Okoye¹

¹*Department of Biochemistry, Faculty of Medical Sciences, University of Jos, Jos, Plateau State, Nigeria.*

²*Department of Chemical Pathology, Federal College of Veterinary and Medical Laboratory Technology, National Veterinary Research Institute, Vom, Plateau State, Nigeria, P.M.B. 02 Vom, Nigeria.*

Authors' contributions

This whole work was carried out in collaboration between both authors. Both authors read and approved the final manuscript.

Original Research Article

Received 15th April 2014
Accepted 25th June 2014
Published 1st August 2014

ABSTRACT

Background and Objective: Epidemiological studies have shown that high body iron stores are associated with insulin resistance and type 2 diabetes. The aim of this study was to evaluate iron status of patients with type 2 diabetes mellitus (T2DM). Blood samples were collected from participants after overnight fast.

Materials and Methods: Two hundred (200) subjects comprising 130 type 2 diabetics attending Hospitals in Jos and 70 normal subjects as controls were involved in the survey. Questionnaires were used in the recruitment of participants. Serum ferritin (SF) was assayed by ELISA method, while other parameters were determined colorimetrically.

Results: Diabetics presented with higher mean age, BMI, and blood pressure than non diabetics. Also, diabetics had elevated serum ferritin, SI, TIBC, total cholesterol (TC), triglycerides, and TC/HDL ratio, lower serum HDL; elevated serum aminotransferases and creatinine than non diabetic subjects. There was a strong and significant positive correlation between serum ferritin levels and each of six diabetes mellitus risk factors: systolic blood pressure, diastolic blood pressure, serum total cholesterol (TC), serum triglyceride (TG), HDL and TC/HDL.

Conclusion: This work has shown that type 2 diabetic subjects exhibited strong positive

*Corresponding author: Email: itspatonline@yahoo.com;

diagnostic features for the indices of iron status, dyslipidaemia, liver damage and kidney dysfunction compared to non diabetic subjects.

Keywords: Ferritin; type 2 diabetes; diabetes risk factors; hyperglycaemia.

1. INTRODUCTION

Type 2 diabetes mellitus (T2DM) is a predominant public health concern worldwide, accounting for 90% of the cases of diabetes globally [1,2]. The chronic hyperglycaemia of diabetes is associated with long-term damage, dysfunction, and failure of various organs, especially the eyes, kidneys, nerves, heart, and blood vessels [3].

In diabetes, lipid abnormalities, anaemia, alteration of liver and kidney functional indices have been implicated as major risk factors to the progression of Diabetic complications [4]. The pathogenesis of type 2 diabetes mellitus (T2DM) is complex and involves the interaction of genetic and environmental factors [5]. Individuals with T2DM show both insulin resistance and beta cell defects [6].

The relationship between T2DM and iron metabolism has gained interest in both research and clinical practice [7]. Several studies indicate that increased body iron stores and subclinical haemochromatosis have been associated with the development of glucose intolerance and type 2 diabetes [8]. Several studies have shown that moderate increases in iron stores below the levels found in patients with hereditary haemochromatosis (HH) were associated with significant elevations in blood glucose and insulin levels [9]. Increased serum ferritin, reflecting body iron overload, is often associated with measures of insulin resistance [8].

Genetic mutations in the haemochromatosis gene (HFE) make up the most common genetic cause of elevated serum ferritin levels [10,11]. Hereditary haemochromatosis (HH) is an autosomal recessive disorder characterized by enhanced intestinal absorption of dietary iron [12]. Defects in human haemochromatosis protein (HFE) cause iron overload due to reduced hepatic hepcidin secretion [13]. Genetic and acquired factors lead to increased iron absorption and storage in hepatocytes, with consequent alteration in the redox status [14].

Excess iron impairs pancreatic β cell function and causes β cell apoptosis [15]. Iron serves as a potent pro-oxidant in human body and participates in the generation of reactive oxygen species (ROS) such as hydroxyl radical [16]. The susceptibility of β -cells to iron-induced oxidative stress and the iron deposition in β -cells usually leads to apoptosis, and consequently, to insulin deficiency [15]. Iron deposition also induces insulin resistance by inhibiting glucose uptake in fat and muscle tissues, and reducing the capacity of liver to extract insulin, which results in an abnormal increase in hepatic glucose production [17].

Although several epidemiological studies have reported a strong association between elevated serum ferritin and increased risk for type 2 diabetes; more so, a link between serum ferritin concentration and insulin resistance or type 2 diabetes has been established [18]. However, it appears that little or no work on the relationship between iron status and type 2 diabetes mellitus has been done in Nigeria, hence, the need for this study.

This study was carried out to evaluate the iron status of patients with type 2 diabetes mellitus (T2DM) in Jos, for the purpose of ascertaining whether the hypothesis regarding the relationship between iron stores and type 2 DM, which has been established by studies in several countries of the world, is also applicable to Nigeria and Plateau state, in particular.

2. MATERIALS AND METHODS

2.1 Study Location/ Sampling Procedures

This study was carried out amongst diabetic patients attending the outpatient medical clinics of Jos University Teaching Hospital, Plateau Specialist Hospital, Jos, and ECWA Evangel Hospitals, Jos and amongst normal subjects as controls. A total of two hundred (200) subjects comprising of one hundred and thirty (130) type 2 DM patients (70 females and 60 males) aged 40-70 years and seventy (70) apparently healthy subjects as controls (37 females and 33 males) were recruited for the study. Patients with acute or chronic inflammatory or infective disease and serious diabetic complications were excluded from the study. Control subjects consist of individuals with no history of medical disorder on the basis of their clinical history and biochemical data.

Ethical clearance from hospital review boards and informed consents from patients/controls were obtained. One standard questionnaire was completed for each subject, which included subject's personal data, drug usage, disease history and physical examination. The designing of questionnaire and recruitment of participants was carried out under a close supervision by a clinician. Confidentiality was maintained in accordance with standard medical practice.

Blood pressure was measured by mercury sphygmomanometer in the right upper arm of the subject, who was seated for 5 minutes before the measurement. Blood pressure was measured twice, and the mean of these two measurements was used in the analysis. Weight was measured without shoes to the nearest 0.1kg on a weighing machine and height was measured to the nearest 0.1m with an anthropometric rod. Body mass index was calculated as weight in kilograms divided by the square of height in metres.

2.2 Sample Collection and Preservation

10ml of venous blood was collected from each volunteer after an overnight fast (2ml into fluoride oxalate bottle for glucose analysis, 2ml into EDTA bottle for some haematological parameters and the remaining into plain tubes for other biochemical analysis. Plasma glucose and haematological analysis were carried out immediately while blood in the plain tube was allowed to clot and centrifuged to separate the serum from the cells for subsequent analysis of other biochemical parameters. Sera were stored frozen at -20°C, prior to assay. Frozen sera were completely thawed and well mixed and all reagents were allowed to attain room temperature.

2.3 Biochemical Analysis

Plasma glucose concentration was determined using glucose oxidase method [19]. Total cholesterol was determined by enzymatic point method [19]. Triglyceride (TG) was determined by enzymatic colorimetric method by hydrolysis of triglycerides [19]. High-density Lipoprotein-cholesterol (HDL-c) was measured colorimetrically in the supernatant fluid by

enzymatic endpoint method (Randox test kit), following the precipitation of other lipoproteins (apo-B-100-containing lipoprotein) with a polyanion-divalent cation mixture (phosphotungstic and magnesium chloride) [20]. Low-density lipoprotein-cholesterol (LDL-c) was calculated by the Friedward equation summarised as follows [21]:

$$\text{LDL} = (\text{Tc} - \text{HDL-c}) - \frac{\text{Triglyceride}}{2.2} \text{ (mmol/L), where Tc = Total cholesterol}$$

Aspartate aminotransferase (AST) and alanine aminotransferase (ALT) were assayed by the kinetic method of Karmen [22].

Serum iron (SI) was assayed by the colorimetric method by Stookey [23] whereas total iron binding capacity (TIBC) was determined by the colorimetric method of Ramsay [24]. Serum ferritin was assayed using human ferritin enzyme immunoassay test kit (catalogue number: 6001) based on the method of White et al. [25].

Serum creatinine was determined by the Jaffes reaction methods [26].

Packed cell volume (PCV) was determined by the micro-haematocrit method [27]. Haemoglobin was assayed by the cyanmethaemoglobin method [27].

White blood cells (WBC) were determined by visual white cell counts [28].

2.4 Statistical Analysis

Statistical analysis was performed with SPSS software version 16.0. Data are expressed as the mean \pm SEM. Independent sample t-test was used for comparison of means of variables between groups while Pearson correlation coefficient was used to determine the relationship between serum ferritin and the outcome variables. Statistical significance was considered when p value was less than 0.05.

3. RESULTS

The results of the physical anthropometric risk indices profiles in diabetic and control subjects are summarised on Tables 1. The mean age of the controls (41.9 \pm 0.5) was significantly ($P=0.000$) lower than that of the diabetics (52.7 \pm 0.8). Expectedly, the mean blood pressure (systolic & diastolic) of control subjects (124.4 \pm 1.5; 79.9 \pm 1.2 in mmHg) were significantly lower than in the diabetics (153.6 \pm 2.3; 91.6 \pm 1.1 in mmHg). There was statistical significant difference between male and female control subjects in height, BMI, and diastolic blood pressure all but serum ferritin being higher in males, while male and female diabetics differ significantly only in height, the mean height being higher in males.

The results of the serum glucose and the indices of iron status in diabetic and control subjects are summarised on Table 2. Expectedly, the mean glucose level was significantly ($P=0.000$) higher in diabetics (9.9 \pm 0.4 mmol/l) than in the control (4.4 \pm 0.1 mmol/l). Similarly, the mean values of the three iron status indices, serum ferritin, serum iron and total iron binding capacity (TIBC), were significantly higher in diabetics than in the control subjects. The differences in the mean PCV and haemoglobin values were not statistically significant ($P=0.431$ and $p=0.401$ respectively). There are statistically significant gender differences between male and female control subjects in respect of serum ferritin, serum iron, PCV and haemoglobin, with the females having the lower values in all four indices.

Table 1. Mean blood pressure and other physical anthropometric indices of DM subjects

Variables	Diabetic			Control			Total		
	Female (n=70)	Male (n=60)	P-value	Female (n=39)	Male (n=33)	P-value	Diabetic (n=130)	Control (n=37)	P value
Age (years)	51.7±1.1	53.8±1.2	.329	42.2±0.7	41.5±0.7	.793	52.7±0.8	41.9±0.5 ^c	.000
Syst BP (mmHg)	151.5±3.2	154.6±3.7	.393	123.3±2.0	125.6± 2.3	.484	153.7±2.3	124.4±1.5 ^c	.000
Diast BP(mmHg)	90.0±1.5	92.7±1.8	.173	80.4±1.4 ^b	79.3±2.1	.028	91.6±1.1	79.9±1.2 ^c	.021
Weight(kg)	69.9±1.9	72.6±1.9	.461	71.9±2.3	69.7±1.9	.225	71.2±1.4	70.8±1.5	.083
Height(m)	1.57±0.01a	1.7±0.0	.808	1.6±0.0 ^b	1.7±0.0	.953	1.6±0.0	1.6 ±0.0	.656
BMI(kg/m)	27.8±0.7	26.2±0.7	.472	29.3± 0.9 ^b	25.9±0.9	.525	27.1±0.5	27.7±0.6	.410

Tabulated values are means ± S.E.M; Statistical significance between means was assessed using independent sample t-test, ^a= depicts significant difference between diabetic female versus diabetic male ($P<0.05$), ^b= depicts significant difference between control female versus control male ($P<0.05$), ^c= depicts significant difference between diabetic (total) and control (total) ($P<0.05$), Abbreviations: syst BP= systolic blood pressure; diast BP=diastolic blood pressure; BMI=body mass index.

Table 2. Mean serum glucose, serum ferritin and other indices of iron status of DM subjects

Variables	Diabetic			Control			Total		
	Female (n=70)	Male (n=60)	P-value	Female (n=39)	Male (n=33)	P-value	Diabetic (n=130)	Control (n=37)	P-value
Glucose (mmol/l)	9.4±0.5	10.5±0.6	.109	4.3±0.1	4.5±0.1	.002	9.9±0.4	4.4±0.1 ^c	.000
Ferritin (ng/ml)	65.8±6.4	86.7±6.6	.471	33.8±4.7 ^b	62.6±7.5	.043	75.7±4.7	47.9±4.5 ^c	.012
SI(μmol/l)	40.8±2.1	46.7±2.2	.944	18.4±0.8 ^b	23.0±1.4	.034	43.5±1.5	20.3±0.8 ^c	.000
TIBC(μmol/l)	87.1±2.5	82.1±2.8	.492	68.3±3.00	67.3±3.1	.701	84.4±1.9	67.9±2.2 ^c	.048
TS (%)	48.1±2.5	58.5±2.6	.823	27.5±1.5 ^b	37.2±3.4	.002	52.9±1.8	31.5±1.7 ^c	.000
PCV (%)	41.4±0.6	42.6±0.7	.749	41.5±0.7b	44.8±0.7	.706	41.9±0.5	43.0±0.5	.431
Hb (g/l)	13.8±0.2	14.1±0.6	.036	13.6±0.2b	14.6±0.2	.366	13.7±0.1	14.0±0.2	.401

Tabulated values are means ± S.E.M; Statistical significance between means was assessed using independent sample t-test, ^a= depicts significant difference between diabetic female versus diabetic male ($P<0.05$), ^b= depicts significant difference between control female versus control male ($P<0.05$), ^c= depicts significant difference between diabetic (total) and control (total) ($P<0.05$), Abbreviations: SI=serum iron; TIBC=total iron binding capacity; TS=transferrin saturation; PCV=packed cell volume; Hb=haemoglobin.

The results of serum lipids and other indicators of clinical complications of type 2 diabetes mellitus in diabetic and control subjects are summarised on Table 3. There are significant differences between the diabetic and control subjects in all four lipid parameters with the mean values for three of these, total cholesterol (TC), triglyceride (TG) and TC-HDL, being higher in diabetics than in the control subjects; the level of the exception, HDL, was lower in diabetics than the control. There are no significant gender differences in these parameters between the female and male subjects either among the diabetics or control subjects. The mean serum creatinine, a muscle tissue and renal function marker, of the diabetics ($108.4 \pm 1.8 \mu\text{mol/l}$) was significantly higher ($P=0.001$) than in the control subjects ($93.4 \pm 1.8 \mu\text{mol/l}$), a situation consistent with a higher state of muscle tissue catabolism or renal dysfunction in diabetics. There is a significant ($P=0.022$) gender difference in serum creatinine between female and male control subjects with the higher value in males.

The activities of the liver marker enzymes, aspartate and alanine aminotransferases, in the serum were significantly elevated in the diabetics as compared to the control subjects, suggesting a hepatotoxic side effect of diabetes in the subjects. There are no significant gender differences.

The results of the analyses of the relationship between serum ferritin levels and each of the other risk factors, physical anthropometric indices, blood glucose, indices of iron status, serum lipid indices and indicators of clinical complication of DM, in all subjects, diabetics and control subjects by Pearson's two-tailed correlation analysis are summarized on Table 4. There is a significant correlation between serum ferritin levels and systolic BP ($P=0.040$) and diastolic BP ($P=0.017$), respectively, among the diabetic subjects. A similar significant relationship obtains in control subjects and when control and diabetics are pooled.

Among diabetic subjects, there is a significant ($P=0.005$) correlation between serum ferritin levels and fasting plasma glucose level. There is no significant correlation between serum ferritin and any of the other iron status indices of DM among the diabetics. However, a significant correlation exists between serum ferritin and serum iron in the control subjects and total sample population.

There is a significant correlation between serum ferritin and each of three serum lipid risk factors, TC, TG and TC/HDL; the correlation with the fourth lipid index, HDL, is also statistically significant. A similar significant correlation between serum ferritin level and each of all but one, HDL exists among the control subjects and when control and diabetic subjects are pooled. No significant correlation exists between serum ferritin and any of markers of cytotoxic side effects, that is, serum aminotransferases (AST, ALT) and serum creatinine.

Table 3. Mean serum concentrations of lipids and other indicators of clinical complications of DM

Variables	Diabetic			Control			Total		
	Female (n=70)	Male (n=60)	P- value	Female (n=39)	Male (n=33)	P- value	Diabetic (n=130)	Control (n=37)	P-value
TC (mmol/l)	5.4±0.2	5.6±0.2	.856	3.9±0.1	4.0±0.2	.404	5.5±0.1	4.00±0.1 ^c	.000
TG (mmol/l)	2.0±0.1	2.1±0.1	.604	1.0±0.1	1.1±0.1	.668	2.1±0.1	1.1±0.1 ^c	.000
HDL (mmol/l)	0.9±0.0	0.8±0.0	.329	1.4±0.1	1.4±0.1	.925	0.9±0.0	1.4±0.1 ^c	.023
TC-HDL	6.4±0.2	6.8±0.2	.608	2.8±0.1	3.0±0.2	.414	6.6±0.2	2.9±0.1 ^c	.000
AST (iu/l)	25.5±1.2	26.0±1.6	.253	22.7±2.2	21.2±1.5	.116	25.7±1.0	22.0±1.3 ^c	.024
ALT (iu/l)	25.9±1.2	27.1±1.6	.126	22.0 ± 1.8	20.1±1.9	.953	26.4±1.0	21.1±1.3 ^c	.001
Creatinine (µmol/l)	107.8±2.6	109.3±2.5	.058	85.6 ± 2.3 ^b	102.8±1.9	.022	108.4±1.8	93.4±1.8 ^c	.001
WBC	4.6±0.1	4.6±0.1	.517	4.6 ± 0.1	4.2±0.1	.167	4.6±0.1	4.6±0.1	.711

Tabulated values are means ± S.E.M; Statistical significance between means was assessed using independent sample t-test, ^a= depicts significance difference between diabetic female versus diabetic male (P<0.05), ^b= depicts significance difference between control female versus control male (P<0.05), ^c= depicts significance difference between diabetic (total) and control (total) (P<0.05)
 Abbreviations: TC=total cholesterol; TG=triglyceride; HDL=high density lipoprotein; AST=aspartate aminotransferase; ALT=alanine aminotransferase; WBC=white blood cell

Table 4. Correlation between serum ferritin and some outcome variables

Variables	All subjects		Diabetics		Control	
	R	P-value	r	P-value	r	P-value
Age (years)	.163*	.028	-.003	.977	.086	.515
Weight (kg)	-.036	.625	-.038	.678	-.052	.693
Height(m)	.119	.111	.155	.085	-.107	.412
BMI (kg/m ²)	.057	.448	.067	.460	.088	.501
Systolic BP (mmHg)	.306**	.000	.185*	.040	.318*	.015
Diastolic BP (mmHg)	.314**	.000	.215*	.017	.315*	.016
Glucose (mmol/l)	.355**	.000	.248**	.005	.872**	.000
SI (μmol/l)	.255**	.001	.128	.181	.131**	.315
TIBC (μmol/l)	.073	.347	.001	.991	-.161	.215
TS (%)	.225**	.003	.104	.275	.200	.122
PCV (%)	.015	.848	.021	.823	.174	.180
Hb (g/l)	.056	.459	.107	.251	.063	.637
TC (mmol/l)	.447**	.000	.368**	.000	.389**	.002
TG (mmol/l)	.375**	.000	.275**	.002	.360**	.004
HDL (mmol/l)	-.148*	.045	.098	.276	-.167	.211
TC/HDL	.422**	.000	.338**	.000	.434**	.001
WBC	.005	.948	-.042	.656	.249	.053
Creatinine(μmol/l)	.089	.237	.025	.781	.133	.308
AST (iu/l)	0.080	.286	.035	.705	.152	.243
ALT (iu/l)	.057	.449	.026	.773	.039	.767

Pearson's correlation coefficient was used to determine the relationship between serum ferritin and the outcome variables, *Correlation is significant at the 0.05 level (2-tailed), **Correlation is significant at the 0.01 level (2-tailed), Abbreviations: BMI=body mass index; BP=blood pressure; SI=serum iron; TIBC=total iron binding capacity; TS=transferrin saturation; PCV=packed cell volume; Hb=haemoglobin; TC=total cholesterol; HDL=high density lipoprotein; WBC=white blood cell; AST=aspartate aminotransferase; ALT=alanine aminotransferase.

4. DISCUSSION

Serum levels of iron store markers, such as ferritin, TIBC and serum iron and the common indicators of clinical complications of type 2 diabetes, such as serum lipid profile, liver enzymes and creatinine, were determined in the present study. Significantly elevated serum ferritin levels in diabetics compared to control subjects recorded in the present study support similar findings from previous studies [29,30,31,32]. The exact mechanism through which elevated ferritin promotes the development of type 2 diabetes is uncertain. Elevated iron stores may induce diabetes through a variety of mechanisms, including oxidative damage to pancreatic beta cells, impairment of hepatic insulin extraction by the liver, and interference with insulin's ability to suppress hepatic glucose production [8]. I.e. iron deposition and iron-induced oxidative stress contribute to the pathogenesis of type 2 diabetes (T2D) through β-cells apoptosis, hepatic dysfunction, and insulin resistance [15]. The pancreatic beta cells are particularly susceptible to oxidative damage because of their weak antioxidant defense [9].

The causative role of elevated iron store levels in the onset of insulin resistance is well established by prospective data as well as evidence that blood donations improve insulin sensitivity by decreasing iron stores [16]. Studies have revealed that first-degree relatives of type 2 diabetes mellitus patients with normal glucose tolerance have higher ferritin

concentrations than normal control subjects [33]. These observations may suggest a genetic predisposition to hyperferritinaemia in type 2 diabetes. Genetic mutations in the haemochromatosis gene (HFE) make up the most common genetic cause of elevated serum ferritin levels [34,35,11]. Hereditary hemochromatosis is an autosomal recessive disorder associated with the HFE genes and is characterized by enhanced intestinal absorption of dietary iron [36,12].

The systemic iron homeostasis is achieved by regulating iron absorption and storage and recycling mechanisms. The ferroportin-mediated efflux of Fe²⁺ from enterocytes and macrophages into the plasma is critical for systemic iron homeostasis [37,38]. This process is negatively regulated by hepcidin, a liver-derived peptide hormone that binds to ferroportin and promotes its phosphorylation, internalization and lysosomal degradation [38,15]. Thus hepcidin acts to decrease the absorption of dietary iron and the release of recycled iron from macrophage stores by diminishing the effective number of iron exporters on the membrane of enterocytes or macrophages [39,40]. Under conditions of high iron, hepcidin-induced down regulation of ferroportin in duodenal enterocytes prevents dietary iron from entering the circulation [15]. Defects in human hemochromatosis protein (HFE) cause iron overload due to reduced hepatic hepcidin secretion [13,41].

Dyslipidaemia in diabetics compared to control subjects observed in this study is in agreement with the findings of other researchers [42,43]. Impaired lipid metabolism resulting from uncontrolled hyperglycaemia has been implicated in diabetes [44,45]. Type 2 DM is associated with a cluster of interrelated plasma lipid and lipoprotein abnormalities that are all recognized as predictors for coronary heart disease [46]. Resistance to insulin likely underlies the changes that occur in lipid parameters of type 2 DM and, usually, it is associated with higher concentrations of TC and TG, and lower concentrations of HDL-C [47,48]. The mechanism responsible for hypertriglyceridaemia may be an increased hepatic secretion of VLDL and a delayed clearance of TG-rich lipoproteins, which might mainly be due to increased levels of substrates for TG production, free fatty acids, and glucose. The latter could be secondary to decreased activity of lipoprotein lipase (LPL), a key enzyme for lipoprotein-TG [48].

The significantly elevated liver marker enzymes in diabetes compared to control subjects also obtained in the present study is in line with the report of previous researchers [49,50]. The occurrence of liver disease and raised liver enzymes is common in type 2 diabetes [51,52]. Elevated serum liver enzymes (aminotransferases and alkaline phosphatase) can reflect abnormalities in liver cells or in the bile duct [53,54]. For example, predominant elevation of aminotransferases typically indicates hepatocellular injury, whereas elevated alkaline phosphatase indicates cholestatic injury. Elevated alkaline phosphatase and aminotransferases can indicate a mixed pattern of injury [54]. AST is present in blood cells and many tissues, including liver, muscle, brain, pancreas, and lung. Although ALT is present in several organs and in muscle, the highest levels are in the liver, making it a more specific indicator of liver disease [54]. Alkaline phosphatase is active in many organs, mainly the liver and bones, but is also found in the small bowel, kidneys, and placenta. Diseases of the hepatobiliary system can cause moderate to marked elevations of alkaline phosphatase [54]. Mild chronic elevations of transaminases often reflect underlying insulin resistance [53].

The elevated serum creatinine level in diabetics compared to control subjects obtained in this study is in accord with the reports of several researchers [55,56]. Renal impairment associated with elevated serum creatinine levels is common in diabetes [57,58]. Diabetic nephropathy occurs in approximately one third of type 2 diabetics [59,60]. Raised plasma

creatinine in diabetic patients may indicate a pre-renal problem such as volume depletion which may be due to impaired function of the nephrons [55]. Gender differences in serum creatinine between female and male control subjects observed in this study may be due to the difference in muscle mass of males and females [56]. An independent role for gender in the progression of renal disease in human has not been clearly established [56]. Since prospective and interventional studies have confirmed an aetiologic role of iron overload in the pathogenesis of insulin resistance and type 2 diabetes, in the future actively lowering body iron stores may become a tool in preventing type 2 diabetes in selected subjects with impaired glucose metabolism.

5. LIMITATION

Serum ferritin concentration is by far the most commonly used indicator of body iron stores in epidemiological studies [61]. However, the specificity of high circulating ferritin levels as a marker of increased body iron stores is somewhat limited because ferritin is an acute-phase reactant that may be elevated in inflammation and other disorders such as liver disease and cancer [62]. In addition, circulating ferritin is also increased with alcohol consumption and body mass index (BMI), and differs with gender [9]. It is thus unclear whether the association of ferritin with type 2 diabetes risk factors reflects these other abnormalities

Serum soluble transferrin receptor (sTfR) concentration has been suggested as a more accurate measure of available body iron [9]. Circulating sTfR levels correlate inversely with body iron stores and thus reflect, inversely, intracellular iron storage. The sTfR:ferritin ratio has been found to distinguish between subjects with similarly high ferritin levels, and sTfR is believed to be free of influence by acute or chronic inflammation, therefore it has been suggested that the sTfR:ferritin ratio is a better marker than ferritin alone to measure a wide range of iron levels to quantifiably reflect body iron over the entire spectrum of iron balance [16]. More studies on the association of iron stores with type 2 diabetes risk need to be carried out more extensively using sTfR:ferritin ratio in addition to serum ferritin as indicators of iron status.

6. CONCLUSION

From the foregoing, a simple blood test which measures ferritin levels can be used to predict development of diabetes in healthy people. This may help in identifying high risk people who would possibly benefit from lifestyle or therapeutic interventions that can lower iron stores in the body.

CONSENT

Informed consent was used in the recruitment of the participants and confidentiality was maintained in accordance with standard medical practice

ETHICAL APPROVAL

Ethical approval was given by the Ethics Committees of Jos University Teaching Hospital, Plateau Specialist Hospital and ECWA Evangel Hospital Jos.

ACKNOWLEDGEMENT

The sponsorship granted POO by the Federal College of Veterinary and Medical Laboratory Technology (FCVMLT), Vom through study leave is duly acknowledged. Study volunteers are also gratefully acknowledged.

COMPETING INTERESTS

Authors have declared no competing interests exit.

REFERENCES

1. Rohilla A, Kumar R, Rohilla S, Kushnoor A. Diabetic retinopathy: origin and complications. *European Journal of Experimental Biology*. 2012;2(1):88-94.
2. Hussain SA, Marouf BH. Flavonoids as alternatives in treatment of type 2 diabetes mellitus. *Academia Journal of Medicinal Plants*. 2013;1(2):031-036.
3. Akyuz F, Tekin N, Aydın O, Temel HE, Isıklı B. The effect of metformin and exercise on serum lipids, nitric oxide synthase and liver nitric oxide levels in streptozotocin-nicotinamide induced diabetic rats. *African Journal of Pharmacy and Pharmacology*. 2012;6(5):336-342.
4. Oyedemi SO, Adewusi EA, Aiyegoro OA, Akinpelu DA. Antidiabetic and haematological effect of aqueous extract of stem bark of *Azelia africana* (Smith) on streptozotocin-induced diabetic Wistar rats. *Asian Pacific Journal of Tropical Biomedicine*. 2011;353-358.
5. American Diabetes Association. Diagnosis and classification of diabetes mellitus. *Diabetes Care*. 2012;35(1):S64-71.
6. Khardori R. Type 2 diabetes mellitus. *Medscape Reference; Drugs, Diseases and Procedures*. Updated 4 April 2012. Accessed 12 April 2012. Available on: <http://emedicine.medscape.com/article/117853>.
7. Zafar Z, Qureshi HJ, Karim A. Insulin resistance and serum parameters of iron status in type 2 diabetics. *Pak J Physiol*. 2011;7(2):28-31.
8. Raj S, Rajan GV. Correlation between elevated serum ferritin and HbA1c in type 2 diabetes mellitus. *International Journal of Research in Medical Sciences*. 2013;1(1):12-15.
9. Bao W, Rong Y, Rong S, Liu L. Dietary iron intake, body iron stores, and the risk of type 2 diabetes: a systematic review and meta-analysis. *British Medical Journal*. 2012;10:119.
10. Adams P. Management of elevated serum ferritin levels. *Gastroenterology and Hepatology*. 2008;4(5):333-334.
11. Li M, Wang L, Wang W, Qi XL, Tang ZY. Mutations in the HFE gene and sporadic amyotrophic lateral sclerosis risk: a meta-analysis of observational studies. *Brazilian Journal of Medical and Biological Research*. 2014;47(3). Available on: <http://dx.doi.org/10.1590/1414-431X20133296>
12. Santos PC, Cançado RD, Pereira AC, Schettert IT, Soares RA, Pagliusi RA, et al. Hereditary hemochromatosis: mutations in genes involved in iron homeostasis in Brazilian patients. *Blood Cells Mol Dis*. 2011;46(4):302-7. doi: 10.1016/j.bcmd.2011.02.008.

13. Bardou-Jacquet E, Philip J, Lorho R, Ropert M, Latournerie M, Houssel-Debry P, et al. Liver transplantation normalizes serum hepcidin level and cures iron metabolism alterations in HFE hemochromatosis. *Hepatology*. 2014;59(3):839-47. doi: 10.1002/hep.26570. Epub 2014 Jan 27.
14. Valenti L, Dongiovanni P, Piperno A, Fracanzani AL, Maggioni M, Rametta R. et al. Alpha1-antitrypsin mutations in NAFLD: high prevalence and association with altered iron metabolism but not with liver damage. *Hepatology*. 2006;44:857-864.
15. Gabrielsen JS, Gao Y, Simcox JA, Huang J, Thorup D, Jones D, et al. Adipocyte iron regulates adiponectin and insulin sensitivity. *Journal of Clinical Investigation*. 2012;122(10):3529–3540
16. Zhao Z, Li S, Liu G, Yan F, Ma X, Huang Z, Tian H. Body iron stores and heme-iron intake in relation to risk of type 2 diabetes: a systematic review and meta-analysis. 2012;PLoS ONE 7(7):e41641. doi:10.1371/journal.pone.0041641.
17. Green A, Basile R, Rumberger JM. Transferrin and iron induce insulin resistance of glucose transport in adipocytes. *Metabolism*. 2006;55:1042–1045.
18. Dongiovanni P, Valenti L, Fracanzani AL, Gatti S, Cairo G, Fargion S. Iron depletion by deferoxamine up-regulates glucose uptake and insulin signalling in hepatoma cells and in rat liver. *American Journal of Pathology*. 2008;172:738-747.
19. Trinder P. Enzymatic colorimetric methods. *Annals of Clinical Biochemistry*. 1969;6:24-27.
20. Bachorik PS, Albers JJ. Precipitation Methods for quantification of lipoprotein. *Methods in Enzymology*. 1986;129:78-100.
21. Friedwald WT, Levy RI, Fredrickson DS. Estimation of concentration of low density lipoprotein cholesterol in plasma, without use of the preparative ultra-centrifuge. *Clinical Chemistry*. 1972;18:499-502.
22. Karmen A. Spectrophotometric assay of glutamic transaminase in serum. *Journal of Clinical Investigation*. 1955;34:131-133.
23. Stookey L. Determination of iron concentration in serum. *Analytical Chemistry*. 1970;42:779.
24. Ramsey WNM. The determination of total iron binding capacity of serum. *Clinical Chemical Acta*. 1957;2:221-226.
25. White D, Kramer D, Johnson G, Hamilton H. Enzymatic immunoassay for the quantitative determination of ferritin concentration in serum. *American Journal of Clinical Pathology*. 1986;72:346.
26. Bonsnes RW, Taussky HH. The colorimetric determination of creatinine by the Jaffe reaction. *Journal of Biological Chemistry*. 1945;158:581-591.
27. Lewis SM, Bain BJ, Bates I. Basic haematological techniques. In: Lewis SM, Bain BJ, Bates I, editors. *Dacie and Lewis Practical Haematology*. 10th ed. Churchill livingstone Elsevier. 2006;25-57.
28. Baker FJ, Silverton RE. *Introduction to Medical Laboratory Technology*. London: Design and Patients by the Copyright Licensing Agency Ltd; 1998.
29. Ford ES, Cogswell ME. Diabetes and serum ferritin concentration among U.S. adults. *Diabetes Care*. 1999;22(12):1978–1983.
30. Sharifi F, Sazandeh SH. Serum ferritin in type 2 diabetes mellitus and its relationship with HbA1c. *Acta Medica Iranica*. 2004;42:142–5.
31. Sharifi F, Zadeh HJ, Nasab NM, Amirmoghaddami H. Serum ferritin concentrations in an impaired fasting glucose population and their normal control group. *Acta Medical Iranic*. 2007;45(3):321–324.
32. Sharifi F, Nasab NM, Zadeh HJ. Elevated serum ferritin concentrations in prediabetic subjects. *Diabetes and Vascular Disease Research*. 2008;5(1):15–18.

33. Ren Y, Tian H, Liang J, Zhao G. Elevated serum ferritin concentration in a glucose impaired population and in normal glucose tolerance first degree relatives in familial type 2 diabetic pedigrees. *Diabetes Care*. 2004;27:622–623.
34. Allen KJ, Bertalli NA, Osborne NJ, Constantine CC, Delatycki MB, Nisselle AE, et al. HFE Cys282Tyr homozygotes with serum ferritin concentrations below 1000 microg/L are at low risk of hemochromatosis. *Hepatology*. 2010;52(3):925-33. doi: 10.1002/hep.23786.
35. Karaca H, Güven K, Önal M, Gürsoy Ş, Başkol M, Özkul Y. The prevalence of primary hereditary hemochromatosis in central Anatolia. *Turk J Gastroenterol*. 2013;24(1):43-50.
36. Phatak P, Brissot P, Wurster M, Adams PC, Bonkovsky HL, Gross J, et al. A phase 1/2, dose-escalation trial of deferasirox for the treatment of iron overload in HFE-related hereditary hemochromatosis. *Hepatology*. 2010;52(5):1671-779. doi: 10.1002/hep.23879.
37. Hentze MW, Muckenthaler MU, Galy B, Camaschella C. Two to tango: regulation of mammalian iron metabolism. *Cell*. 2010;142(1):24-38.
38. Ganz T. Heparin and iron regulation, 10 years later. *Blood*. 2011;117(17):4425-4433.
39. Finberg KE. Unraveling mechanisms regulating systemic iron Homeostasis. *Haematology*. 2011;532-537.
40. Fleming RE, Ponka P. Iron Overload in Human Disease. *New England Journal of Medicine*. 2012;366:348-59.
41. Adams PC. Heparin in hemochromatosis: the message or the messenger? *Hepatology*. 2014;59(3):749-50. doi: 10.1002/hep.26715. Epub 2014 Jan 13.
42. Samatha P, Venkateswarlu M, Siva Prabodh V. Lipid profile levels in type 2 diabetes mellitus from the tribal population of Adilabad in Andhra Pradesh, India. *Journal of Clinical and Diagnostic Research*. 2012;6(4):590-592.
43. Sarkar BC, Saha HR, Azad AK, Sana NK, Choudhury S. Serum lipid profile status of type 2 diabetic patients in the cross section population in Dhaka City of Bangladesh. *Bangladesh Journal of Medical Science*. 2012;11(2):121-125.
44. Uttra KM, Devrajani BR, Shah SZA, Devrajani T, Das T, Raza S, Naseem M. Lipid profile of patients with diabetes mellitus (A multidisciplinary study). *World Applied Sciences Journal*. 2011;12(9):1382-1384.
45. Emeka EN, Ebele JI, Ignatius CM, Osita A, Ibegbu MD, Miriam OA, et al. Lipid Profile and glycated haemoglobin level In type-2 diabetic patients in Enugu, South-East Nigeria. *Journal of Asian Scientific Research*. 2012;2(5):300-306.
46. Gordon L, Ragoobirsingh D, Morrison YA, Choo-Kang E, McGrowder D, Martorell E. Lipid Profile of type 2 diabetic and hypertensive patients in the Jamaican Population. *Journal of Laboratory Physicians*. 2010;2(1):25–30.
47. Shaikh MA, Kumar S, Ghouri RA. Type 2 diabetes mellitus and lipid abnormalities. *Journal of Liaquat University of Medical and Health Sciences*. 2010;9(3):145-147.
48. Kumawat M, Singh I, Singh N, Singh V, Kharb S. Lipid peroxidation and lipid profile in type 2 diabetes mellitus. *Diabetes*. 2012;3(3):3147.
49. Sarkar BC, Saha HR, Sarker PK, Sana NK, Sayeed MA, Choudhury S. Liver enzymes in diabetic and non diabetic subjects with clinically diagnosed hepatitis. *Ibrahim Medical College Journal*. 2011;5(2):46-50.
50. Morling JR, Strachan MW, Hayes PC, Butcher I, Frier BM, Reynolds RM, et al. Prevalence of abnormal plasma liver enzymes in older people with type 2 diabetes. *Diabetic Medicine*. 2012;29(4):488-91.
51. Forlani G, Di Bonito P, Mannucci E, Capaldo B, Genovese S, Orrasch M. Prevalence of elevated liver enzymes in Type 2 diabetes mellitus and its association with the metabolic syndrome. *Journal of Endocrinological Investigation*. 2008;31(2):146-52.

52. Villegas R, Xiang Y, Elasy T, Cai Q, Xu W, Li H. Liver enzymes, type 2 diabetes, and metabolic syndrome in middle-aged, urban Chinese men. *Metabolic Syndrome and Related Disorders*. 2011;9(4):305–311.
53. Elizabeth H, Harris MD. Elevated liver function tests in type 2 diabetes. *Clinical Diabetes*. 2005;23:115-119.
54. Aragon G, Younossi ZM. When and how to evaluate mildly elevated liver enzymes in apparently healthy patients. *Cleveland Clinic Journal of Medicine*. 2007;77(3):195-204.
55. Aldler AI, Stevens RJ, Manley SE, Bilous RW, Cull CA, Holman RR. Development and progression of nephropathy in type 2 diabetes: The United Kingdom Prospective Diabetes Study. *Kidney International*. 2003;63:225-232.
56. Wagle TJ. Gender wise comparison of serum creatinine and blood sugar levels in type 2 diabetic patients. *Bombay Hospital Journal*. 2010;52:64-68.
57. Erejuwa OO, Sulaiman SA, Wahab MS, Sirajudeen KN, Salleh MS, Gurtu S. Glibenclamide or metformin combined with honey improves glycemic control in streptozotocin-induced diabetic rats. *International Journal of Biological Science*. 2011;7:244-252.
58. Cao J, Li C, Zhang P, Cao X, Huang T, Bai Y, Chen K. Antidiabetic effect of burdock (*Arctium lappa* L.) root ethanolic extract on streptozotocin-induced diabetic rats. *African Journal of Biotechnology*. 2012;11(37):9079-9085.
59. Rehman G, Khan SA, Hamayun M. Studies on diabetic nephropathy and secondary diseases in type 2 diabetes. *International Journal of Diabetes in Developing Countries*. 2005;25:25-29.
60. Idonije BO, Festus O, Oluba OM. Plasma glucose, creatinine and urea levels in type 2 diabetic patients attending a Nigerian Teaching Hospital. *Research Journal of Medical Sciences*. 2011;5(1):1-3.
61. Montonen J, Boeing H, Steffen A, Lehmann R, Fritsche A, Joost HG, et al. Body iron stores and risk of type 2 diabetes: results from the European Prospective Investigation into Cancer and Nutrition (EPIC)-Potsdam study. *Diabetologia*. 2012;55(10):2613–2621.
62. Rajpathak SN, Crandall JP, Wylie-Rosett J, Kabat GC, Rohan TE, Hu FB. The role of iron in type 2 diabetes in humans. *Biochim Biophys Acta*. 2009;1790(7):671-681.

© 2014 Okonkwo and Okoye; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/3.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:

The peer review history for this paper can be accessed here:
<http://www.sciencedomain.org/review-history.php?iid=597&id=19&aid=5611>