

Journal of Complementary and Alternative Medical Research

Volume 20, Issue 3, Page 18-24, 2022; Article no.JOCAMR.92768 ISSN: 2456-6276

Evaluation of Antidiarrheal Activity of Aqueous Extract of *Cucurbita moschata* (Pumpkin) Leaf in Castor Oil Induced Diarrhea Wistar Rats

Maryam Usman Ahmed ^{a*}, Isaac John Umaru ^b, Abraham Henry Haruna ^a and Diowato Titus ^a

^a Department of Biochemistry, Adamawa State University, Mubi, Adamawa, Nigeria.
^b Department of Biochemistry, Federal University, Wukari, Taraba State, Nigeria.

Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/JOCAMR/2022/v20i3416

Open Peer Review History:

This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here: https://www.sdiarticle5.com/review-history/92768

Original Research Article

Received: 18/08/2022 Accepted: 21/10/2022 Published: 27/12/2022

ABSTRACT

Background: Diarrhea is one of the major health problems in developing countries leading to mortality and morbidity among children under 5 years of age. This study evaluated the antidiarrheal activities of *Cucurbita moschata* on castor oil-induced diarrheal wistar rats.

Methods: Thirty (30) wistar rats were divided into six groups of five rats each. All rats except group 1 received 1mL castor oil to induce diarrhea. Groups I and II served as the normal and negative control. Group III received the standard drug (loperamide), groups IV-VI were treated with 100, 200 and 300 mg/kg b.wt of aqueous leaf extract of *Cucurbita moschata* respectively. Stool inhibition, castor oil-induced enteropooling, and gastrointestinal motility test were determined to evaluate the antidiarrheal effect of the extract.

^{*}Corresponding author: E-mail: maryamahmed566@gmail.com;

J. Compl. Altern. Med. Res., vol. 20, no. 3, pp. 18-24, 2022

Results: Percentage stool inhibition significantly (p < 0.05) increased with increase in the dose of the extract. The percentage distance travelled by the charcoal of the groups treated with the plant extract significantly decreased when compared to the negative control. This decrease was comparable with the group administered loperamide. Administration of aqueous extract of *Cucurbita moschata* leaf significantly decreased the volume of the intestinal fluid when compared with the volume of intestinal fluid of the negative control and the group administered loperamide. **Conclusion:** Aqueous leaf extract of *Cucurbita moschata* possess antidiarrheal activity against castor oil-induced diarrhea.

Keywords: Diarrhea; stool inhibition; gastrointestinal motility; enteropooling; Cucurbita moschata.

1. INTRODUCTION

Diarrhea а gastrointestinal disorder is characterized by changes in bowel movement, frequency, and changes in the consistency of feces with increased water content, making it liquid or pasty, which can manifest acutely or chronically [1]. Diarrhea is one of the major health problems worldwide especially in developing countries for children under 5 years of age [2]. It occurs mainly due to poor hygiene practices, lack of improved sanitation facilities and low hygienic status of shared sanitation facilities [3,4]. Diarrhea etiology is associated with a wide array of putative pathogens such as rotavirus, Cryptosporidium, E. coli, Shigella, Adenovirus, Aeromonas, V. cholera, C. jejuni, Norovirus, Rotavirus. Salmonella enterica. Sapovirus. Astrovirus and Clostridium difficile with E. coli being the most common among infants [5,6,7]. Enteropathogenic E. coli (EPEC) continues to be the most important cause of diarrhea in children under 2 years of age while enterotoxic E. coli (ETEC) is the most common in children over 2 years [8]. Diarrheal disease is classified based on its duration either as acute (1-13 days), persistent (14 days and above), or chronic (above 30 days), or based on the physiological mechanisms as secretory, inflammatory, osmotic, and motor, where the majority of the etiologies possess complex pathophysiology involving one or more of these mechanisms [9,10,11]. The principles of managing diarrhea diseases are correct diagnosis and treatment of the specific agent that caused the diarrhea [12]. Oral rehydration therapy (ORT) containing electrolytes are administered to prevent or treat dehydration caused by persistent diarrhea, and medications such as antimotility agents like loperamide, agents, antimicrobial antibiotics, and nitazoxanide an anti-parasitic agent are used for the treatment of diarrheal disease when an accurate diagnosis is done and the causative agent is known [13]. However, a recent report

establishes that loperamide induces toxic cardiac arrhythmias and death when taken in high doses [14]. The side effects of other antidiarrheal drugs include nausea, drug toxicity, dizziness, dependency, sedation, respiratory depression, and constipation. Hence, there is need to search for new antidiarrheal drug with fewer side effects. Cucurbita moschata (pumpkin) is a major and important annual dicotvledonous vegetable crop from the family Cucurbitaceae, extensively cultivated worldwide [15]. It grows up to 5 m with creeping and climbing stems bearing tendrils. The stems and leaves are fairly hairy. The stems are strong and cylindrical or perpendicular with petioles measuring 12-30 cm while the leaves are circular, having a kidney shape, heart shape, or triangular shape. The flowers grow up to 12 cm long, are bell-shaped and largely yellow [16]. Pumpkins can be cultivated in warm areas all over the world due to their cheap growth and production of high nutrient content [17,18]. Due to the rich nutritional composition of Cucurbita moschata such as carbohydrates, flavonoids, phenolics, vitamins, and amino acids [19], it is believed to have functional and health benefits [20]. Thus, it has been proven to possess various medicinal properties that include wound healing [21], antidiabetic, anti-carcinogenic, antioxidant [22], antibacterial [23], antidepressant, andanteinflammatory properties without notable ulcerogenic effect often associated with antiinflammatory drugs [24]. Traditionally, the plant is used to treat diarrhea. There is no scientific report on the antidiarrheal activity of the leaves of Cucurbita moschata. Therefore, this study evaluated the antidiarrheal effects of Cucurbita moschata on diarrheal wistar rats.

2. MATERIALS AND METHODS

2.1 Collection of Plant Material

Fresh matured leaves of *Cucurbita moschata* were collected in May 2022 and authenticated at the Dept. of Botany, Adamawa State University,

Mubi. It was washed and air-dried under shade at room temperature and then pulverized using mortar and pestle into powder form. The powdered sample was stored in a well-tight container and kept at room temperature until required.

2.2 Extraction of the Plant Material

The powdered plant material was extracted using the maceration method described by Azwanida [25]. The plant material was soaked in distilled water in the ratio of 1:4 in a stoppered container and allowed to stand at room temperature with frequent agitation for 3 days. The mixture was pressed and strained by filtration using Whatman filter paper no. 1 after 3 days and the filtrate was evaporated to dryness using a crucible and water bath at 40^oC.

2.3 Experimental Animals

Adult albino rats with a weight range of 150 – 180 g were purchased from the Animal Resource Unit, National Veterinary Research Institute (NVRI) VOM, Plateau State, Nigeria and were housed in wired cages well ventilated, which were allowed free access to drinking water and fed with standard laboratory diet. Guidelines for the protection and handling of laboratory animals by the International Council for Laboratory Animal Science (ICLAS) rats were used in handling the rats. Animals were allowed to acclimatize to the laboratory environment for one week before the experiment commenced.

2.4 Stool Inhibition

Six groups of five animals each were fasted for 12 hours and thereafter castor oil at a dose of 1 mL/rat was administered to rats in groups II - VI to induce diarrhea using an orogastric cannula. Thirty (30) minutes after castor oil administration, rats of group II (negative control) received 1.0 mL of 0.9% NaCl in distilled water (normal saline), and group III received 2 mg/kg b. wt. loperamide (standard drug), groups IV-VI received 100, 200, and 300 mg/kg b.wt. of aqueous leaf extract of Cucurbita moschata, p.o. respectively. The animals were placed separately in metabolic cages over white clean Whatman filter paper, which was changed every hour. The severity of diarrhea was assessed each hour for 4 hours. The total number of diarrhea feces in the control group was considered 100%.

% inhibition = (Control - Test) × 100/Control.

2.5 Measurement of Gastrointestinal Transit

Six groups of five animals each were fasted for 12 hours and thereafter castor oil (1 mL) was administered orally to the rats in groups II - VI. One hour later, group III received the standard drug loperamide (2 mg/kg p.o) while rats of groups IV-VI received 100, 200, and 300 mg/kg b.wt. of leaf extract of Cucurbita moschata, p.o. respectively. After 30 min of the administration, 1 mL of charcoal meal, (10% suspension in 5% gum acacia) was orally administered to rats in each group. The rats were sacrificed by ether (20% v/v) anesthesia and the small intestine was carefully separated from mesentery to avoid being stretched. For each animal, gastrointestinal transit was calculated as the percentage distance traveled by charcoal meal to the total length of the intestine. The inhibitory effect of the extracts on gastrointestinal transit was calculated relative to the respective group.

2.6 Castor Oil-induced Enteropooling

Thirty adult albino rats were randomly divided into six groups. Castor oil (1 mL) was administered orally to rats in groups II - VI. One hour later, group III received the standard drug, loperamide (2 mg/kg p.o.). Rats of groups IV-VI received 100, 200, and 300 mg/kg b.wt. of aqueous leaf extract of Cucurbita moschata, p.o. respectively. After 2 hours of treatment, the rats were sacrificed by ether anesthesia. The edges of the intestine from the pylorus to the caecum were tied with thread and the intestine was removed and weighed. Intestinal fluid was milked into a graduated tube, and the intestinal fluid volume was taken. The intestine was reweighed and differences between full and empty intestines were calculated.

2.7 Statistical Analysis

The mean and statistical analysis computation was done using SPSS software version 24.0. Data are expressed as the mean \pm S.D for a group of five animals. It was statistically analyzed with one-way analysis of variance (ANOVA) and Duncan Multiple Range Test (DMRT). For all the tests, results with p values < 0.05 was considered significant.

3. RESULTS AND DISCUSSION

Table 1 shows the effect of aqueous leaf extract of *Cucurbita moschata* on castor oil-induced

diarrhea. The plant extract significantly (p < 0.05) decreased the number of wet stool when compared with the standard drug, loperamide, and the negative control. The percentage inhibition of the treatment group significantly (p < 0.05) increased as the dose of the extract increased. The percentage stool inhibition of the group treated with the extract significantly (p <0.05) increased when compared with the group treated with the standard drug and the negative control. Induction of diarrhea by castor oil is due to the most active component, ricinoleic acid, which causes inflammation and irritation of the intestinal mucosa leading to the release of prostaglandins which contributes the to pathophysiological functions in the gastrointestinal tract resulting in the stimulation of secretion by increasing the intestinal volume contents and prevention of re-absorption of water [26,27,28]. "Loperamide which acts by increasing phasic segmenting the colonic activities through inhibition of the presvnaptic cholinergic nerves in the submucosal and myenteric plexus was used as a positive control in this study" [29]. "These effects of loperamide results in the reduction of the postprandial flow of and absolute net colonic digesta water absorption, while the relative digesta flow remains unchanged or is transiently reduced thereby reducing the frequency of defecation" [30].

The percentage inhibition of the castor oilinduced diarrhea by the leaf extract of the *Cucurbita moschata* may indicate that the extract is an effective inhibitor of diarrhea. The observed decrease in the number of stools by the plant extract indicates the antidiarrheal potential of the extract. Comparing the results of the groups treated with the extract and the group treated with the standard drug suggests that the plant extract is an effective agent for inhibition of diarrhea than the synthetic drug, loperamide.

Table 2 shows the percentage distance travelled by charcoal. "All the treated groups including the group treated with the standard drug were not significantly (p < 0.05) different from each other however, there was a significant (p < 0.05) decrease when compared with the negative control group. This significant (p < 0.05) decrease observed in the gastrointestinal transit by the extract may indicate that the extract has antimotility activity responsible for decreasing the peristaltic movement and secretion" [31,32]. "It may also indicate that the plant extract possess anticholinergic compounds since castor oil has been suggested to be indirectly mediated by the cholinergic system" [33].

Table 3 shows the effects of *Cucurbita moschita* leaf extract on castor oil-induced enteropooling. "When compared to the negative control group, the volume of intestinal fluid of all the treated groups significantly (p < 0.05) decreased. The volume of intestinal fluid of the group that received 300 mg/kg b.wt. of the extract decreased significantly from the other treated groups. The significant reduction observed in the intestinal fluid volume, especially at a higher dose of the extract suggests that the extract has an inhibitory effect on ricinoleic action. It may also be due to the fact that the extract enhances water reabsorption by decreasing the intestinal

Table 1. Effect of aqueous leaf extract of <i>Cucurbita moschata</i> on			
castor oil-induced diarrhea			

Groups	Wet stool	% Stool inhibition
Gloups		
Group I (Normal control)	0 ± 0.00^{a}	100 ± 0.00^{e}
Group II (Negative control)	12 ± 0.15 ^e	0 ± 0.00^{a}
Group III (2 mg/kg loperamide)	8 ± 0.23^{d}	49 ± 0.12^{b}
Group IV (100 mg/kg b.wt. extract)	$5 \pm 0.09^{\circ}$	$72 \pm 0.09^{\circ}$
Group V (200 mg/kg b.wt. extract)	3 ± 0.13^{b}	83 ± 0.95^{d}
Group VI (300 mg/kg b.wt. Extract)	2 ± 0.15^{b}	90 ± 1.23^{d}

Values are presented as mean \pm S.D. n=5.

Values with different superscript down the column are significantly different at (n < 0.05)

Table 2. Effect of aqueous leaf extract of Cucurbita moschata on castor oil induced gastrointestinal motility

Groups	% Distance travelled by the charcoal		
Group I (Normal control)	30.51 ± 4.01^{a}		
Group II (Negative control)	$46.53 \pm 2.74^{\circ}$		
Group III (2 mg/kg loperamide)	35.43 ± 3.70^{b}		
Group IV (100 mg/kg b.wt. extract)	$38.90 \pm 3.57^{\circ}$		
Group V (200 mg/kg b.wt. extract)	34.24 ± 4.31^{b}		
Group VI (300 mg/kg b.wt. Extract)	40.31 ± 2.23^{b}		

Values are presented as mean \pm S.D. n = 5. Values with different superscript down the column are significantly different at (p < 0.05)

Table 3. Effect of aqueous leaf extract of Cucurbita moschata on castor oil-induced enteropooling

Groups	Volume of intestinal fluid	
Group I (Normal control)	1.46 ± 0.15^{ab}	
Group II (Negative control)	$2.42 \pm 0.27^{\circ}$	
Group III (2 mg/kg loperamide)	1.84 ± 0.12^{b}	
Group IV (100 mg/kg b.wt. extract)	1.54 ± 0.06^{ab}	
Group V (200 mg/kg b.wt. extract)	1.55 ± 0.12^{ab}	
Group VI (300 mg/kg b.wt. Extract)	1.20 ± 0.04^{a}	

Values are presented as mean \pm S.D. n = 5. Values with different superscript down the column are significantly different at (p < 0.05)

motility to allow for proper reabsorption" [34]. The antienteropooling effect of the extract may indicate that the extract is potent in phytochemicals such as steroids that are capable of stimulating the Na⁺ absorption by stimulating any of the apical transporters and reducing mucosal secretion [35]. The antidiarrheal activity of *Cucurbita moschata* observed in this study may indicate the presence of phytochemicals in the extract since earlier studies revealed that bioactive components such as tannins, saponins, flavonoids, alkaloids, sterols, and reducing sugar from medicinal plants possess anti-diarrheal property [31,36,37].

4. CONCLUSION

The results revealed that *Cucurbita moschata* aqueous leaf extract reduced the number of wet stools. It has antimotility and antienteropooling effects. It is more efficient than loperamide. The leaf extract has antidiarrheal potential.

CONSENT

It is not applicable.

ETHICAL APPROVAL

Animal Ethic committee approval has been collected and preserved by the author(s).

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- Smieja M, Goldfarb DM. Molecular detection of diarrheal pathogens. Clin. Microbiol. Newsl. 2016;38(2016):137–145.
- Asrie AB, Abdelwahab M, Shewamene Z, Gelayee DA, Adinew GM, Birru EM. Antidiarrheal activity of methanolic extract of the root bark of *Cordia africana*. J. of Experiment. Phamacol. 2016;8:53–59.
- 3. Semba RD, Kraemer K, Sun K, De Pee S, Akhter N, Moench-Pfanner R, Bloem MW. Relationship of the presence of a household improved latrine with diarrhea and under-five child mortality in Indonesia. The Ame. J. of Trop. Med. and Hygiene. 2011;84(3):443.

 Simiyu S, Swilling M, Cairncross S, Rheingans R. Determinants of quality of shared sanitation facilities in informal settlements: case study of Kisumu, Kenya. BMC Pub. Health. 2017;17(1):1-13.

 Nataro JP, Mai V, Johnson J, Blackwelder WC, Heimer R, Tirrell S, Hirshon JM. Diarrheagenic escherichia coli infection in Baltimore, Maryland, and New haven, Connecticut. Clin. Infect. Dis. 2006;43(4): 402-407.

- Kotloff KL, Nataro JP, Blackwelder WC, Nasrin D, Farag TH, Panchalingam S, Levine MM. Burden and aetiology of diarrhoeal disease in infants and young children in developing countries (the Global Enteric Multicenter Study, GEMS): a prospective, case-control study. The Lancet. 2013;382(9888):209 - 222.
- Qadri F, Svennerholm AM, Faruque ASG, Sack RB. Enterotoxigenic Escherichia coli in developing countries: epidemiology, microbiology, clinical features, treatment, and prevention. Clin. Microbiol. Rev. 2005;18(3):465-483.
- Nguyen TV, Le Van P, Le Huy C, Gia KN, Weintraub A. Etiology and epidemiology of diarrhea in children in Hanoi, Vietnam. Inter. J. of Infect. Dis. 2006; 10(4):298 - 308.
- 9. Dantas RO, Diarreia E. Constipação Intestinal. Medicina. 2004; 37 (2004): 262– 266.
- 10. Patel, K., Thillainayagam, A.V., Diarrhoea. Med.. 2009;37(2009):23 – 27.
- 11. Bhutta Z, Syed S. Diarrheal Diseases. 2016;361–372.
- 12. Camilleri M, Sellin JH, Barrett KE. Pathophysiology, evaluation, and management of chronic watery diarrhea. Gastroenter. 2017;152(3):515 - 532.
- 13. DuPont HL. Persistent diarrhea: a clinical review. Jama. 2016;315(24):2712-2723.
- Dierksen J, Gonsoulin M, Walterscheid JP. Poor man's methadone: A case report of loperamide toxicity. The Ame. J. of Forensic Med. and Path. 2015; 36(4):268-270.
- 15. Wu T, Luo S, Wang R, Zhong Y, Xu X, Lin Y, He X, Sun B, Huang H. The first Illumina-based de novo transcriptome sequencing and analysis of pumpkin (*Cucurbita moschata* Duch.) and SSR marker development. Mol Breeding; 2014.

DOI 10.1007/s11032-014-0128-x

 Marie-Magdeleine C, Mahieu M, Archimède H. Pumpkin (*Cucurbita* moschata Duchesne ex Poir.) Seeds as an Anthelmintic Agent? In Nuts and seeds in health and disease prevention (pp. 933-939). Academic Press.Kazuo KOIKE, Wei LI, Lijuan LIU, Emiko HATA, and Tamotsu NIKAIDO. New Phenolic Glycosides from the Seeds of *Cucurbita moschata*. Chem. Pharm. Bull. 2011;53(2):225–228.

- Kumar S, Rattan P, Samnotra R. Squashes and gourds. In Pessarakli, M. (Ed.) Handbook of cucurbits: growth, cultural practices, and physiology. Boca Raton, FL: CRC Press. 2016;513 -531. ISBN-13 978-1-4822-3459-6.
- Provesi JG, Amante ER. Carotenoids in pumpkin and impact of processing treatments and storage. Processing and Impact on Active Components in Food. 2015;71-80. ISBN-13 978- 0-12-404699-3.
- Wang P, Liu JC, Zhao QY. Studies on nutrient composition and utilization of pumpkin fruit. J. Inner Mongolia Agric. Univ. 2002;23:52–54.
- 20. Zhang F, Jiang ZM, Zhang EM. Pumpkin function properties and application in food industry. Sci. Technol. Food Indus. 2000;21:62–64.
- Bahramsoltani R, Farzaei MH, Abdolghaffari AH, Rahimi R, Samadi N, Heidari M, Amin G. Evaluation of phytochemicals, antioxidant and burn wound healing activities of *Cucurbita moschata* Duchesne fruit peel. Iranian J. of Basic Med. Sci. 2017;20(7):798.
- 22. Yadav M, Jain S, Tomar R, Prasad GBKS, Yadav H. Medicinal and biological potential of pumpkin: an updated review. Nut. Res. Rev.. 2010;23(2):184-190.
- 23. El Zawane Kamarudin, Q. U. A., Helaluddin, A. B. M., Sirajudin, Z. N. M., Chowdhury, A. J. K. Studies on bactericidal efficacy of pumpkin (*Cucurbita moschata* Duchesne) peel. 2014; 2(2): 146 -153.
- 24. Eleiwa NZ, Bakr RO, Mohamed SA. Phytochemical and pharmacological screening of seeds and fruits pulp of *Cucurbita moschata* Duchesne cultivated in Egypt. Inter. J. of Pharmacog. and Phytochem. 2014;29(1):1226 - 1236.
- 25. Azwanida NN. A Review on the Extraction Methods Use in Medicinal Plants, Principle, Strength, and Limitation. Medical and Aromatic Plants. 2015;4:196. DOI:10.4172/2167-0412.1000196.
- Ammon HV, Thomas PJ, Phillips SF. Effects of oleic and ricinoleic acids on net jejunal water and electrolyte movement. Perfusion studies in man. The J. of Clin. Inves. 1974;53(2):374-379.
- 27. Luderer JR, Dermers LM, Nomides C, Hayes AH. Mechanism of castor oil: A biochemical link to the prostaglandins. In:

Samuelson B, Ramwell PW, Paoletti R. (eds). Advances in Prostagladin and Thromboxane Research, Raven Press, New York. 1980;8:1633 - 1635.

- 28. Nwachoko N, Jack IR. Phytochemical screening and antidiarrhoeal activities of *Tetracarpidium conophorum* induce in albino rats. Sky J of Biochem. Res. 2015;4(4):021–024.
- 29. Okere OS, Sangodele JO, Tade OG, Obafemi OT, Falode JA. Anti-diarrhea potential and acute toxicity studies of methanolic extract of *Vernonia amygdalina* and *Cymbopogon citratus* against castor oil induced diarrhea model in rats. Inter. *J.* of Biochem. Res.and Rev. 2015;6(2):46.
- Theodorou V, Fioramonti J, Hachet T, Bueno L. Absorptive and motor components of the antidiarrhoeal action of loperamide: an *In vivo* study in pigs. Gut. 1991;32(11):1355-1359.
- Galvez J, Zarzuelo A, Crespo ME, Lorente MD, Ocete MA, Jimenez J. Antidiarrhoeic activity of *Euphorbia hirta* extract and isolation of an active flavonoid constituent. Planta Medica. 1993;59(04):333-336.
- 32. Brunton LL. Agents affecting gastrointestinal water flux and motility; emesis and antiemetics; bile acids and pancreatic enzymes. Goodman and Gilman's: the pharmacological basis of therapeutics. 1996;917-936.

- Brown JH, Taylor P. Cholinergic agonist In Brunton LL, Lazo JS. Parker KL. (Eds). The pharmacological basis of Therapeutics. 11th edn. McGraw-Hill. New York. 2006;183-200.
- 34. Maiti A, Dewanjee S, Mandal SC. *In vivo* evaluation of antidiarrhoeal activity of the seed of Swietenia macrophylla King (Meliaceae). Trop. J. of Pharm. Res. 2007;6(2):711-716.
- 35. Ahmed MU, Arise RO, Umaru IJ. Identification and biochemical characterization of anti-enteropooling compounds from *Annona senegalensis* root bark. Scientific Afr. 2022;15(2022): e0128.
- 36. Galvez J, Zarzuelo A, Crespo ME, Utrilla MP, Jimenez J, Spiessens C, De Witte P. Antidiarrhoeic activity of *Sclerocarya birrea* bark extract and its active tannin constituent in rats. Phytotherapy Res. 1991;5(6):276-278.
- 37. Otshudi AL, Vercruysse A, Foriers A. Contribution to the ethnobotanical. pharmacological phytochemical and studies of traditionally used medicinal plants in the treatment of dysentery diarrhoea Lomela and in area, Democratic Republic of Congo (DRC). J. of Ethnopharmacol. 2000;71(3):411-423.

© 2022 Ahmed et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history: The peer review history for this paper can be accessed here: https://www.sdiarticle5.com/review-history/92768