



Fatty Acid Compositions of Mixed Microalgae from Tilapia Fish Ponds

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Authors' contributions

This work was carried out in collaboration between both authors. Author NO designed the study, performed the statistical analysis, wrote the protocol, and wrote the first draft of the manuscript. Authors HO managed the analyses of the study. Both authors read and approved the final manuscript.

Article Information

DOI: 10.9734/AJOB/2019/v8i130055

Editor(s):

(1) Dr. Emmanuel Ikechukwu Nnamonu, Lecturer, Department of Biology, Federal College of Education, Eha-Amufu, Nigeria.

Reviewers:

(1) Aliyu Bhar Kisabo, Nigeria.

(2) Dr. K. D. Mini, Mahatma Gandhi University, India

Complete Peer review History: <http://www.sdiarticle3.com/review-history/49284>

Original Research Article

Received 03 March 2019

Accepted 23 May 2019

Published 29 May 2019

ABSTRACT

Microalgae has been getting broad attention of researchers and investors lately, especially when discussing on healthy food and energy sources for the future. In this study, twelve samples of mixed microalgae from outdoor ponds were analyzed for their fatty acid compositions. The potential of microalgae to solve variety of world's problems was not realized because of bottleneck in microalgal supplies at reasonable cost. Therefore, the objective of this study is to determine fatty acid profiles of mixed microalgae from tilapia fish ponds. The study was conducted in Tapak Ternakan Ikan, Taman Pertanian Universiti and Department of Biology, Faculty of Science, Universiti Putra Malaysia. Mixed microalgae were extracted for their lipids with methanol: chloroform mixture and after transesterification, the fatty acid methyl ester were analyzed using gas chromatography equipped with flame ionization detector. Results showed that saturated was the major constituent fatty acids. The average percentages of saturated fatty acids, monounsaturated fatty acids, and polyunsaturated fatty acids obtained were $45.62 \pm 1.37\%$, $20.05 \pm 1.14\%$, and $34.33 \pm 3.17\%$ respectively. The most dominant fatty acid profiles were C18:3n3 (α -linolenic acid) and C16:0 (palmitic acid), with the overall percentages of 19.97% and 19.40% respectively. The fatty acid profiles of mixed microalgae was good with a decent balance of saturated, monounsaturated and polyunsaturated fatty acids.

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Keywords: Mixed microalgae; lipids; fatty acids; gas chromatography.

1. INTRODUCTION

Microalgae have become an alternative, with great efficiency of renewable green source, thanks to the ability of microalgae to convert solar energy into many useful chemical compounds at a faster rate. Studies have shown that microalgae offer promising potential in providing world population for food [1], feed [2], fuel [3] and other essential chemicals [4]. Most of that past and present studies of microalgae focused more on finding the potential species with high productivity and lipid content [5,6,7] culture conditions and culture systems [8,9]. One aspect of microalgae that is of interest lately is lipid content and fatty acid profiles. This is because microalgae can produce lipid many folds compared to terrestrial plant. Fatty acids are one of the metabolites produced by microalgae, that enrich their utility both in the form of food and fuels. Microalgae are widely used as a suitable source of fatty acids traditionally for many years [10,11]. The accumulation of fatty acids by microalgae is well studied and discussed by many researchers [7,12]. Among of the researches that have been carried out are on the compositions of fatty acids in *Spirulina platensis* [13] and the triglycerol content in microalgae [14]. Most research has been focusing on the use of microalgal oils containing long-chain polyunsaturated fatty acid as nutraceuticals products. Omega-3 fatty acids are commonly processed from fish oil. But, due to decreasing of fish oil supplies in recent years, also because of the unpleasant taste and odour of fish oils as well as poor oxidative stability, the potential of fish oils is less promising [15]. Compared to oils from fish, microalgae are considered as self-producing omega-3 and the process is straightforward and economical.

Lipids from microalgae are also being studied as feedstock for the sustainable supply of biodiesel [2,16]. The choice of feedstock for biodiesel production has developed from the edible vegetable oils to the non-edible oils, and now emphasizes on microalgae. The interest on the potential use of microalgae as feedstock for biodiesel production has been increasing in recent years. Dependency on petroleum-based fuels is not sustainable because of the increase of fuel price, lessening of crude oil supply as well as the environmental impact of fossil fuel

usage [17]. Earlier studies demonstrated that under selected conditions, microalgae have potential to produce oil for biodiesel 40 times compared to the oil seed crops per unit land area [18]. But, in the scarcity of publicly available data, it is still unclear whether such gains can be realized for a mass commercial scale production. Hence, the economically potential of microalgal based biofuels to significantly affect present and future needs remains in doubt. To study the practicality of microalgae oil as biodiesel substitute, it is necessary to study the fatty acid profile as only lipid with certain carbon chain are suitable for biodiesel conversion. The properties of biodiesel are mainly determined by the structure of its component fatty acid esters. Among the range of the fatty acids found, the saturated medium-chain fatty acids (C8 to C14) are ideal for biodiesel [19]. On the other hand, biodiesel from PUFAs showed good cold-flow characteristics but it is particularly susceptible to oxidation [14].

In recent years, fatty acids compositions of microalgae in large scale for commercial production have created interests among researchers. Most of the present and past studies on microalgal lipids are done in laboratory conditions using single species. The potential of lipid production in microalgae cells is species-specific and this can also be applied to the ability to produce PUFA [20]. Efforts are being focused on the establishment of optimal conditions for mass production of microalgae with high quality of lipid content. Large-scale of microalgae propagation activity in Malaysia is very limited compared to its market potential and its wide use for both domestic and international market. Among the targeted products from the algal resources in Malaysia is microalgal lipids, especially polyunsaturated fatty acid and carotenoids [21]. To make the process of producing microalgae in large-scale available, the production has to depend on free sunlight available, atmospheric carbon dioxide and nutrients present in wastewater. The highcost of microalgal culture systems relates to the need for a stable culture strain and a practical system. The system not only must be productive, but also exhibit cost and energy efficient. Among the new prospects, there are possibility to produce microalgal biomass in large-scale outdoor production by the application of mixed microalgae to substitute

the monospecies microalgae farming that prone to crash. In this regard, we try to determine the potential of mixed microalgae cultured in tilapia pond to replace the practice of monoculture and high-cost photobioreactor system. The principal objective was to determine the fatty acid compositions of mixed microalgae from tilapia fish ponds.

2. MATERIALS AND METHODS

2.1 Mixed Microalgae Samples Collection

Twelve samples of mixed microalgae were collected from different tilapia fish ponds located in Taman Pertanian Universiti, Universiti Putra Malaysia (UPM). The mixed microalgae were harvested using flocculation agent, ferric chloride (FeCl_3). Then, the collected samples were filtered using nylon clothes, washed three times, and kept in -80°C freezer for at least three days. After that, the samples were freeze-dried and ground into powder for further analysis.

2.2 Extraction of Lipid

In this study, the Folch method [22] was employed to extract the lipid from mixed microalgae samples. The powder of mixed microalgae samples was weighed and homogenized with chloroform:methanol (2:1 by volume) to a final volume 20 times the volume of mixed microalgae samples (1 g in 10 ml of solvent mixture). After that, the mixtures were sonicated for five minutes in ice bath before the whole mixtures were agitated for 24 hours at room temperature in an orbital shaker. Next, the liquid phase was recovered by centrifuging at 5000 rpm for 30 minutes. After centrifuged, the bottom layer was discarded. The recovered solvent was washed with 0.9% sodium chloride (NaCl) solutions. The mixtures were centrifuged again at 2000 rpm for 30 minutes to separate the phases. The upper layer was discarded by siphoning and the lower layer was evaporated under vacuum using rotary evaporator at 40°C .

2.3 Fatty Acid Methyl Ester Preparation

0.5 g of samples oils were weighed and transferred into a vial with a tight sealing cap using a pasteur pipette. One milliliter of hexane was added into the vial and vortexed briefly to dissolve the lipids. 0.5 ml of sodium methoxide

was added and vortexed again for one minute. After that, the clear upper layer of methyl ester was pipetted off prior to analysis.

2.4 Gas Chromatography Analysis

The analyses were performed on a SHIMADZU GC2010 equipped with a flame ionization detector (FID) system. Helium was utilized as the carrier gas. To assist in the confirming identification, the standard Supelco 37-component FAME Mix (47885-U) contained methyl esters of fatty acids ranging from C4 to C24 was used. Based on the chromatogram, the compositions of fatty acid profile were evaluated by comparing the retention time of each peak and its area with standard.

3. RESULTS AND DISCUSSION

Commercial productions of microalgae in open pond system have been fully established by different microalgae ventures to produce valuable products from microalgal biomass. The main advantages of open systems are the lower capital costs together with low maintenance and reduced energy needs [23]. Most microalgal cultivation nowadays are grown as monocultures. The main reason for this is due to the specific strain of microalgae containing the high value by-product desired for harvest. The main drawback of monocultures in open systems is they are prone to contamination since cultures are exposed to the environment directly [24]. For this reason, mixed culture microalgae were used to substitute the monocultures cultivation. A culture of mixed microalgae composed of various strains with various optimal growing conditions would be less susceptible to environmental shifts compared to a monoculture. In this study, the samples of lipids from mixed microalgae collected from tilapia fish ponds were extracted to determine the fatty acid compositions.

From the study, the fatty acid compositions varied among the samples, with relatively high percentage of saturated fatty acid ($45.62 \pm 1.37\%$). The second major constituent was polyunsaturated fatty acid with the percentage of $34.33 \pm 3.17\%$ and the least percentage was monounsaturated fatty acid with the percentage of $20.05 \pm 1.14\%$. Fig. 1 showed the average percentage of fatty acid compositions in 12 samples of mixed microalgae obtained from this study.

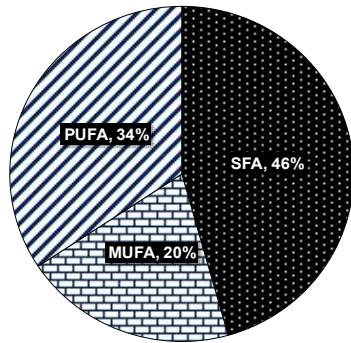


Fig. 1. Overall percentage of fatty acid compositions in mixed microalgae obtained from present study

The total cellular composition of fatty acid, the lipid class and the length of fatty acid chain as well as the degree of saturation are highly varied among microalgae species and the culture conditions. As shown in Fig. 2, the saturated fatty acid (SFA) content were in contrary with polyunsaturated fatty acid (PUFA) content. This might be due to the differences in the culture conditions, growth phase and also the variation of microalgae available in the culture.

Table 1 represented the five most prominent fatty acids of mixed microalgae identified in this study. From previous studies, the most

common fatty acid profiles of microalgae were palmitic (C16:0), stearic (C18:0), oleic (C18:1), linoleic (C18:2) and linolenic acids (C18:3) [25, 26]. From present study, all of the dominant fatty acids were comparable to [25]. These results also suggested that mixed microalgae culture produces fatty acid compositions comparable to those of pure microalgal species in many literatures [27,28]. Previously, a study was conducted on biodiesel production using mixed microalgal culture grown in domestic wastewater [9]. In the study, the palmitic (C16:0), palmitoleic (C16:1), stearic (C18:0), oleic (C18:1), linoleic (C18:2) and linolenic (C18:3) acid methyl esters were found to be the predominant fatty acid constituents. All of these fatty acids were also predominantly found in present study. Most of the microalgae investigated by many literatures have comparable fatty acid profile, but the proportion of fatty acid for each microalgae is different. This is largely depending on the strain used and culture conditions [29].

As proposed by a study [11], microalgae are considered as a good source of polyunsaturated fatty acids and the data collected from present study agreed with the statement. From 29 classes of fatty acid compositions determined in this study, the polyunsaturated fatty acid, α -linolenic acid (C18:3n3) was the most abundant in mixed

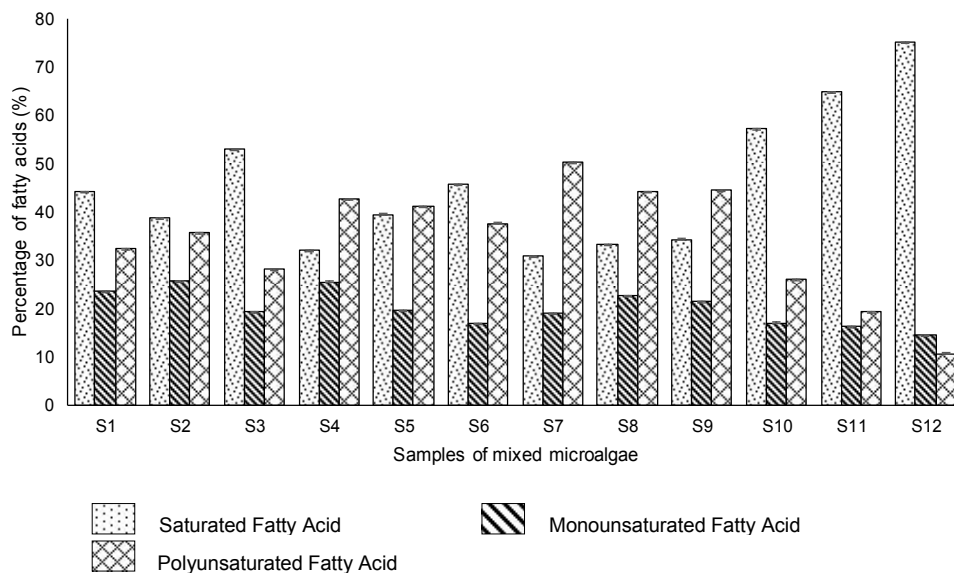


Fig. 2. Percentage of fatty acid compositions in mixed microalgae in twelve different tilapia ponds obtained from present study

Table 1. The major fatty acids composition found in the mixed microalgae samples in this study

Fatty acid	Common name	Average percentage (%)
C18:3n3	α -linolenic acid	19.97
C16:0	Palmitic acid	19.40
C18:2n6	Linoleic acid	13.80
C18:1n9	Oleic acid	8.42
C11:0	Undecylenic acid	8.30

Table 2. Comparisons of fatty acid composition in several vegetables oil and microalgae oils

	SAF	GRA	SUN	WHE	PUM	SES	ALM	RAP	COC	NAN	ISO	MIX	MIX	MIX
SFA	9.3	10.4	9.4	18.2	19.6	16.9	9.3	6.3	92.1	37.1	20.4	43.7	55	46
MUFA	11.6	14.8	28.3	20.9	26.1	42	67.9	72.8	6.2	22.8	17.0	32.4	35.3	20
PUFA	79.1	74.9	62.4	61	54.3	41.2	22.8	20.9	1.6	37.8	39.9	23.9	9.7	34
Omega-3	0.2	0.2	0.2	1.2	0.1	0.2	0	1.2	0	uk	uk	uk	2.1	20.1
Omega-6	79	74.7	62.2	59.7	54.2	40.9	22.8	19.6	1.6	uk	uk	uk	7.6	14.1
	[30]									[31]		[9]	[32]	Present study

Data are expressed as percentages of total fatty acid methyl esters (FAMES); uk means that FAs was unknown. Abbreviations of the samples mean: SAF- Safflower, GRA- Grape, SUN- Sunflower, WHE- Wheat germ, PUM- Pumpkin seed, SES- Sesame, ALM- Almond, RAP- Rapeseed, COC- Coconut oils, NAN- Nannochloropsis oceania, ISO- Isochrysis galbana, MIX- Mixed microalgae

Microalgae in this study. In present study, α -linolenic acid was the most abundant and present in all samples of mixed microalgae. The third major fatty acids determined in this study was also polyunsaturated fatty acid which is linoleic acid (C18:2n6). α -linolenic acid is an omega-3 type of PUFA while linoleic acid is an omega-6 polyunsaturated fatty acid. This suggested that mixed microalgae mass production have potential to be developed as the main source of polyunsaturated fatty acid mainly omega-3 to substitute the present source of omega-3 that mainly comes from fish. The high content of palmitic acid of mixed microalgae found in this study make it possible to be utilized in the production of biodiesel.

4. CONCLUSION

In present study, twelve samples of mixed microalgae collected from tilapia ponds were used to extract lipids and determine the fatty acid compositions. The study revealed that the compositions of fatty acid from mixed microalgae exhibited balance proportion of saturated and unsaturated fatty acids. The mixed microalgae cultivated in outdoor production have a comparable fatty acid compositions as high-maintenance monoculture microalgae. Besides, in view of economics and practicality of the production of biomass, the mixed microalgae cultivation has a very high potential as substitute for current monoculture system. This result can be improved by optimizing few parameters depend on targeted by-products.

ACKNOWLEDGEMENTS

The authors acknowledge the Taman Pertanian Universiti, UPM for providing the place for microalgal biomass collection and access to their facilities.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

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