



***In-vitro* Antioxidant Screening of Ethanol Extracts of *Costus afer* and *Justicia carnea* Leaves**

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Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

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ABSTRACT

This study evaluated the *in-vitro* antioxidant activity of ethanol extract of *Costus afer* and *Justicia carnea* leaves. Ethanol extract of the plant leaves were obtained using standard procedures. The antioxidant parameters of the plants extract studied were 2,2-diphenyl-1-picrylhydrazyl (DPPH) radical, Ferric Reducing Antioxidant Power (FRAP) and Nitric oxide. In each case, the result of test was compared with that of a standard Ascorbic Acid (Vitamin C). The antioxidant study showed that *Justicia carnea* extract had a significantly higher Nitric oxide radical scavenging ability (04.16 ± 0.68) compared with *Costus afer* (02.15 ± 0.26) at 50 $\mu\text{g/ml}$ concentration while DPPH scavenging ability of the extract of *Justicia carnea* showed no significant difference with *Costus afer* (07.63 ± 0.42) and (06.29 ± 0.53) at 50 $\mu\text{g/ml}$ concentrations respectively. For FRAP test, the result indicated that ethanol extract of *Costus afer* (01.78 ± 0.22) was significantly lower ($p < 0.05$) than that of *Justicia carnea* (05.12 ± 0.22) at 800 $\mu\text{g/ml}$ concentration. The findings show that ethanol extracts of *Costus afer* and *Justicia carnea* could operate as main antioxidants and free radical scavengers. The findings back up local assertions that they can be used to treat malaria in folklore medicine.

Keywords: Antioxidants; *Costus afer*; *Justicia carnea*; antioxidant; medicinal plants.

1. INTRODUCTION

“Plants and its derived natural products have received considerable attentions recently

because of its pharmacological properties such as antioxidant activity” [1]. “Medicinal herbs have therefore become of interest due to their prospects in meeting the health needs of

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mankind.” [2,3]. “Among all components used in battling chronic diseases, phytochemicals, plant-derived molecules endowed with steady antioxidant power have been at the forefront. The cumulative and synergistic activities of the bioactive molecules present in plant food have been reported to be responsible for their enhanced antioxidant properties” [4].

“Oxidative damage to proteins and nucleic acids gives rise to a variety of specific damaged products as a result of modifications of amino acids or nucleotides” [5]. “Such oxidative damage might lead to cellular dysfunction, and it is this that might contribute to the pathophysiology of a wide variety of diseases”.

“In Nigeria and many African countries, herbs and leafy vegetables are used as food, food drinks, and medicinal purposes” [6]. “The use of herbs requires good knowledge of the toxicity, dosage purity, and suitable extraction solvent and adverse effects for effective usage” [7,8].

“ It is known as “Okpete” in Igbo, in Hausa “tete-egun” “Kakizawa” in Yoruba, and “Mbriem” in Efik all in Nigeria.” [9]. “This plant is used in the treatment of inflammation, arthritis, asa laxative, purgative, diuretic, in rheumatism, and treatment of several other diseases” [10]. “Synthetic drugs are used in the treatment of disease but because of the high cost and side effects associated with their use [11], attention is now directed towards the use of medicinal plant products in the prevention or management of most diseases”.

“*Justicia carnea* (Flamingo plant) is a flowering plant of *Justicia*, belonging to the *Acanthaceae* family” [12]. “It is widely distributed in various parts of Africa. In Nigeria, the shrubs are grown around homesteads and act as fences. *Justicia carnea* is called “hospital too far” in some parts of Nigeria while others refer to it as “ogwu obara” meaning blood tonic. Traditionally, several species of *Justicia* are used in the management of inflammation, gastrointestinal disorders, respiratory tract infection, fever, pain, diabetes, diarrhea, liver diseases, rheumatism, and arthritis” [13,12]. “Phytochemical analysis of leaves crude extracts of *Costus afer* and *Justicia carnea* has revealed the presence of flavonoids, saponins, alkaloids, tannins, phenols, and glycosides” [14,15].

This study aims to determine the scientific bases for the use of *costus afer* and *Justicia carnea* by evaluating their *in-vitro* antioxidant activities. This

is significant because, particularly in developing countries, there is a growing desire for medical plants and plant products as alternatives to orthodox medicines.

2. MATERIALS AND METHODS

2.1 Plant Collection

The leaves of *C. afer* were harvested at Ihiagwa, Owerri West Local Government Area, Imo State, while leaves of *J. carnea* were harvested at Umuezeala, Eziobodo in Owerri West Local Government Area, Imo state Nigeria.

2.2 Plant Identification

The fresh leaves were identified by Prof. D. I. Edet of the Department of Forestry and Wildlife Technology, School of Agriculture and Agricultural Technology (SAAT), FUTO. The plants were authenticated by another taxonomist Dr. F. A. Faruwa of the Department of forestry and wildlife technology, SAAT, FUTO. The leaves of *C. afer* were prepared and kept at the herbarium with voucher number FUTO/FWT/HERB/2019/056, and for *J.carnea* FUTO/FWT/HERB/2019/057.

2.3 Plant Extraction

The plant leaves were harvested in large quantities and then thoroughly washed to get rid of unwanted particles before being air-dried at room temperature (27 °C- 31 °C) for about one (1) month to constant weight under shade. The dried samples were pulverized into the powdered form using a diesel-powered grinder and then stored separately in an air-tight containment. A quantity of 300g of each powdered sample was soaked separately in 1800ml of absolute ethanol of analytical grade, for 72 hours. Each sample solution was filtered. The filtrates were separately concentrated using a water-bath at a temperature of 45 °C. All extracts were weighed and then stored in well-stoppered containers and preserved in a refrigerator maintained at a temperature of 4 °C until subsequent use.

2.4 Laboratory Analysis

The antioxidant parameters of the plant extracts analyzed by spectrophotometric methods were 2,2-diphenyl-1-picrylhydrazyl(DPPH) radical, Ferric Reducing Antioxidant Power (FRAP), and Nitric oxide. In each case result of the test was

compared with that of a standard Ascorbic Acid (Vitamin C).

2.5 DPPH Radical Scavenging Activity Assay

“DPPH radical scavenging activity of the samples was estimated as described” by Mensor et al., 2001. The crude extract at concentrations (50,100, 200, 400, and 800) µg/ml each was mixed with 1ml of 0.5 mM DPPH (in ethanol) in a cuvette. The absorbance at 517 nm was taken after 30 minutes of incubation in the dark at room temperature. The experiment was done in triplicate. The percentage of antioxidant activities were calculated as follows:

$$\% \text{ Antioxidant activity (AA)} = 100 - \left[\frac{\text{Sample} - \text{Blank}}{\text{control}} \times 100 \right]$$

One milliliter of methanol plus 2.0 ml of the test extract was used as the blank while 1.0 ml of the 0.5 mM DPPH solution plus 2.0 ml of methanol was used as the negative control. Ascorbic acid (Vitamin C) was used as the reference standard.

2.6 Reducing Power Assay

The reducing property of the samples was determined as described by Benzie and Strain (1996). FRAP working solution was prepared by mixing Acetate buffer (300 mM) at pH 3.6 (3.1 g sodium acetate, 3H₂O and 16ml glacial acetic acid in 1000 ml buffer solution) as solution 1, and then 2, 4, 6-triphridyl-s-triazine (TPTZ) (10 mM) in 40 mM HCL as solution 2 and finally FeCl₃ 6H₂O (20 mM) in distilled water as solution 3.

FRAP working solutions were prepared by mixing solution 1, 2, and 3 in the ratio of 10:1:1, respectively. The working solutions were freshly prepared for each test. For calibration, an

aqueous solution containing a known quantity of ascorbic acid was utilized. FRAP reagent; blank Sample; 50, 100, 200, 400, and 800 µg/ml concentrations of FRAP reagent (3ml) and 100 µl sample solution were mixed and left to stand for 4 minutes. At 37°C, coulometric values were taken at 593 nm. A parallel procedure was used to test the ascorbic acid standard solution. A calibration curve was used to make the calculations.

2.7 Nitric Oxide Scavenging Activity Assay

Nitric oxide reacts with oxygen in an aqueous solution at physiological pH to form nitrite ions, which can be quantified using the Griess reaction.

The reaction mixture (3ml) containing sodium nitroprusside (10mM) in phosphate buffer saline (PBS) and the extract from (50 – 800) µg/ml were incubated at 25°C for 15 minutes. After incubation, 0.5 ml of the reaction mixture was removed and 0.5ml of Griess reagent (1% (w/v) sulfanilamide, 2% (w/v) H₃PO₄ and 0.1% (w/v) naphthyl ethylenediamine hydrochloride) was added. The absorbance of the chromophore formed was measured at 546 nm.

2.8 Statistical Analysis

Statistical analysis was carried out with the aid of IBM SPSS statistics for windows; SPSS Inc., Chicago, Standard version 20 to determine differences between the mean of the tests. Post-hoc analysis was also performed to deduce the level of significant differences between the variables. All analyses were performed in triplicate. Data obtained was analyzed using multiple analysis of variance (MANOVA) and the results were expressed as mean ± standard deviation. $P < 0.05$ was considered significant.

3. RESULTS

Table 1. Percentage (%) inhibition toward DPPH free radicals

Concentration (µg/ml)	Activities of <i>J.carnea</i>	Activities of <i>C. afer</i>	Activities of Ascorbic acid
50	07.63 ± 0.42 ^a	06.29 ± 0.53 ^a	42.02 ± 0.45 ^b
100	48.93 ± 0.86 ^b	21.91 ± 1.57 ^a	68.94 ± 0.74 ^c
200	52.88 ± 0.56 ^b	29.95 ± 0.72 ^a	80.16 ± 1.68 ^c
400	74.58 ± 0.89 ^b	60.26 ± 1.63 ^a	83.18 ± 0.91 ^c
800	87.93 ± 0.87 ^b	80.38 ± 0.88 ^a	92.61 ± 0.58 ^c

The analysis was carried out in triplicates and the results were presented as mean ± standard deviation. The rows bearing different superscripts are statistically different at $p < 0.05$

Table 2. Percentage (%) reducing power of *Justicia carnea* and *Costus afer* FRAP

Concentration (µg/ml)	Activities of <i>J. carnea</i>	Activities of <i>C. afer</i>	Activities of Ascorbic acid
50	00.10 ± 0.01 ^b	00.00 ± 0.00 ^a	00.39 ± 0.02 ^c
100	00.44 ± 0.03 ^b	00.04 ± 0.03 ^a	00.63 ± 0.01 ^c
200	01.35 ± 0.07 ^b	00.54 ± 0.04 ^a	03.53 ± 0.13 ^c
400	02.12 ± 0.18 ^b	00.92 ± 0.03 ^a	04.58 ± 0.42 ^c
800	05.12 ± 0.22 ^b	01.78 ± 0.02 ^a	06.94 ± 0.24 ^c

The analysis was carried out in triplicates and the results were presented as mean ± standard deviation. The rows bearing different superscripts are statistically different at $p < 0.05$

Table 3. Percentage (%) inhibition toward nitric oxide radicals

Concentration (µg/ml)	Activities of <i>J. carnea</i>	Activities of <i>C. afer</i>	Activities of Ascorbic acid
50	04.16 ± 0.68 ^b	02.15 ± 0.26 ^a	05.91 ± 0.12 ^b
100	21.31 ± 1.29 ^b	04.82 ± 0.17 ^a	25.26 ± 0.46 ^c
200	36.11 ± 1.12 ^b	19.29 ± 0.73 ^a	60.89 ± 0.63 ^c
400	46.69 ± 0.61 ^b	31.91 ± 1.74 ^a	79.21 ± 0.76 ^c
800	79.04 ± 1.30 ^b	50.47 ± 1.23 ^a	90.16 ± 0.53 ^c

The analysis was carried out in triplicates and the results were presented as mean ± standard deviation. The rows bearing different superscripts are statistically different at $p < 0.05$

4. DISCUSSION

“The medicinal value of plants lies in some chemical substances that produce definite physiological action on the human body” [16]. “There is ample evidence to support the health benefits of medicinal plants” [17]. “Because plants contain complex mixtures of bioactive compounds, information on the potential health of individual phytochemical is linked to information on the health effects of plants that contain those phytochemicals” [18].

“These phytochemicals are known to possess a variety of biological activities including antimicrobial, antioxidant, anti-inflammatory and anticancer activities” [19].

“Several research works have been carried out on countless plants and they have received great attention because they contain high amounts of known antioxidants such as polyphenols, vitamin C, etc. The consumption of these plants has been reported to be inversely associated with morbidity and mortality from degenerative diseases” [20]. Another study by Jack et al. [21] reported that “the reducing power of plants correlates with its phenolic content”.

There might be possible toxic effect of chronic usage of these extracts. The results of the study by Kalu et. al., 2020 suggested that chronic administration of ethanol extract of *Ficus*

capensis at 250 mg/kg and 150 mg/kg may induce some level of anemia and may lead to infection, due to the presence of contaminants in such leaves at a higher dosage.

Table 1 shows the results of the samples' radical scavenging activity against the stable 2, 2-diphenyl-1-picrylhydrazyl (DPPH) free radical as compared to the reference antioxidant ascorbic acid. It clearly shows that as concentration increases from 50 µg/ml to 800 µg/ml, samples exhibited increasing scavenging activity for both *Costus afer* and *Justicia carnea*. results further revealed the highest scavenging activity of 87.93 ± 0.87 % at a concentration of 800 µg/ml for *Justicia carnea*, while *Costus afer* showed the highest scavenging activity of 80.38 ± 0.88% at the same concentration.

“It has been reported that *Costus afer* and *Justicia carnea* are rich sources of flavonoids and other phenolics” [22,23,12]. “The cumulative and synergistic activities of the bioactive molecules present in medicinal plants have also been reported to be responsible for their enhanced antioxidant properties” [24].

“DPPH radical is known to be used as the model system to investigate the scavenging activities of most natural compounds” [25]. “DPPH is scavenged by antioxidants through the donation of proton forming the reduced DPPH which can be quantified by the decreased absorbance” [26].

“The high DPPH scavenging activity of the plant extracts recorded in this study would be attributed to the high phytochemical constituents. This is in line with various studies on the scavenging abilities of flavonoids” [27], (Zhang et al., 2012)

“The reducing power of any compound can be used as an indicator of its ability to serve as an antioxidant” [28], “it acts by donating hydrogen that subsequently stabilizes free radicals” [29]. The present study indicated varying reducing capacity that trail that of the DPPH scavenging assay. *Justicia carnea* showed the best antioxidant property with regards to reducing power of $05.12 \pm 0.22\%$ at a concentration of 800 $\mu\text{g/ml}$, while *Costus afer* did not indicate any noticeable antioxidant property with regards to reducing power at 50 $\mu\text{g/ml}$. The consistent high value of *Justicia carnea* and its closeness to the values of ascorbic acid which was used as a reference antioxidant suggests that it has the best antioxidant property with regards to reducing power [30,31]. This property can be attributed to the presence of important biopharmaceutical phytochemicals [32,33].

Nitric Oxide is involved in the mediation of important physiological activities like the regulation of cellular toxicity [34]. The result indicated that nitric oxide scavenging activity varies with concentration. As concentration increases from 50 $\mu\text{g/ml}$ to 800 $\mu\text{g/ml}$, samples exhibit increasing activity for both *Costus afer* and *Justicia carnea*. *Justicia carnea* showed the highest scavenging activity of $79.04 \pm 1.30\%$ at a concentration of 800 $\mu\text{g/ml}$, while *Costus afer* showed the lowest scavenging activity of $02.15 \pm 0.26\%$.

5. CONCLUSION

This study serves as scientific proof of the use of *Justicia carnea* and *Costus afer* in folklore medicine for treatment of various sicknesses. The leave extracts exhibited significant pharmacological activities that serve as a link to its antioxidant capability. However, more research on the potential toxicity of long-term use is required.

DISCLAIMER

The products used for this research are commonly and predominantly used products in our area of research and country. There is no conflict of interest between the authors and

producers of the products because we do not intend to use these products as an avenue for any litigation but the advancement of knowledge. Also, the research was not funded by the producing company rather it was funded by the personal efforts of the authors

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Karthikumar S, Vigneswari K, Jegatheesan K. Screening of antibacterial and antioxidant activities of leaves *Eclipta prostrata* (L). Scientific Research and Essays. 2007;2(4):101-104.
2. Ameh SJ, Obodozie OO, Inyang SU, Abubakar SM, Garba M. Current phytotherapy; A perspective on the science and regulation of herbal medicine. Journal of Medicinal Plants Research. 2010;4:72-81.
3. Ige SF, Akhigbe RE, Olaleye SB, Adeyemi JW. Gastroprotective of the methanolic extract of *Garcinia kola* in rats. International Journal of Medicine and Biomedical Research. 2012;1:172-178
4. Abdalla AE. The role of antioxidant (Vitamin E) in the control of lead pollution and enhancement of growth within Nile tilapia (*Oreochromis niloticus*). International Journal of Applied Research Veterinary Medicine. 2009;3: 97-101
5. Sataro G, Zsolt R. Implications of oxidative damage to proteins and DNA in aging and its intervention by caloric restriction and exercise. Journal of Sport and Health Science. 2013;2(2):75-80
6. Nwaogu LA, Alisi CS, Ibegbulem CO, Igwe CU. Phytochemical and antimicrobial activity of ethanolic extract of *Landophia Oweriensis* Leaf. African Journal Biotechnology. 2007;6(7): 890-893.
7. Paulo A, Duarte A, Gomes ET. *In vitro* antibacterial screening of *Cryptolepis anguinolenta* alkaloids. Journal of Ethnopharmacol. 1994;44:127-130. Available: [https://doi.org/10.1016/0378-8741\(94\)90079-5](https://doi.org/10.1016/0378-8741(94)90079-5)
8. Murray A. Dietary reference intake for antioxidant nutrients. Journal of the academy of nutrition and dietetics. 1998; 100:637-640.

9. Oliver B. Ibadan: Nigerian College of Arts, Sci and Tech, University Press Nigeria; Medicinal Plants in Nigeria. 1960:1–33.
10. Awouters F, Niemegeers CJ, Lenaerts FM, Janssen PA. Delay of castor oil diarrhoea in rats: A new way to evaluate inhibitors of prostaglandin biosynthesis. *Journal of Pharmacy and Pharmacology*. 1978;30(1): 41–45.
Available: <https://doi.org/10.1111/j.2042-7158.1978.tb13150.x>.
11. Chattopadhyay RR, Bandyopadhyay M. Effect of *Azadirachta indica* leaf extract on serum lipid profile changes in normal and streptozotocin induced diabetic rats. *African Journal of Biomedical Research*. 2005;8:101–104.
12. Correa GM, Alcantara A. Chemical constituents and biological activities of species of *Justicia*: A review. *Brazilian Journal of Pharmacognosy*. 2012;22(1):220-238.
Available: <http://dx.doi.org/10.1590/S0102-695X2011005000196>
13. Badami S, Aneesh R, Sankar S, Sathishkumar MN, Suresh B, Rajan S. Antifertility activity of *Derris brevipes* variety coriacea. *Journal Ethnopharmacology*. 2003;84:99–104.
PMID: 12499083
Available: [https://doi.org/10.1016/s0378-8741\(02\)00288-x](https://doi.org/10.1016/s0378-8741(02)00288-x)
14. Iwu MM. Traditional Igbo Medicine. Applied Sciences. Institute of Africa Studies Publication, University of Nigeria, Nsukka. 2009;3(4):21-25.
15. Anaga AO, Njoku CJ, Ekejiuba ES, Esiaka MN, Asuzu IU. Investigations of the methanolic leaf extract of *Costus afer*. Ker for pharmacological activities *in vitro* and *in vivo*. *Phytomedicine* 2004;11(2-3):242–248.
Available: <https://doi.org/10.1078/0944-7113-00349>
16. Hussain W, Badshah L, Ullah M. Quantitative study of medicinal plants used by communities residing in Koh-e-Safaid Range, northern Pakistani-Afghan borders. *Journal of Ethnobiology and Ethnomedicine* 2018;14:30.
Available: <https://doi.org/10.1186/s13002-018-0229-4>
17. Awuchi CG. Medicinal plants: The medical, food, and nutritional Biochemistry and uses. *International Journal of Advanced Academic Research Sciences, Technology and Engineering*. 2019;5(11).
18. Ahmad SU, Shuid AN, Isa NM. Antioxidant and anti-inflammatory activities of *Marantodes pumilum* (blume) kuntze and their relationship with the phytochemical content. *Records of Natural Products*. 2018;12(6):518.
Available:<https://doi.org/10.25135/RNP.58.17.11.188>
19. Ndukwe GI, Clark PD, Jack IR. *In vitro* antioxidant and antimicrobial potentials of three extracts of *Amaranthus hybridus* L. leaf and their phytochemicals. *European Chemical Bulletin*. 2020;9(7): 164-173.
20. Rodriguez QA, Costa HS. Analysis of carotenoids in vegetable and plasma sample: A review. *Journal of food composition and analysis*. 2006;19: 97-111
21. Jack IR, Clark PD, Ndukwe GI. Evaluation of phytochemical, antimicrobial and antioxidant capacities of *Pennisetum purpureum* (Schumach) extracts. *Chemical Science International Journal*. 2020;29(4):1-14.
22. Anyasor GN, Odunsanya K, Ibeneme A. Hepatoprotective and *in vivo* antioxidant activity of *Costus afer* leaf extract against acetaminophen-induced hepatotoxicity in rats. *Journal of Investigative Biochemistry*. 2013;2:53–61.
Available:<https://doi.org/10.5455/jib.20130301030851>
23. Ukpabi CF, Agbafor KN, Ndukwe OK, Agwu A, Nwachukwu SN. Phytochemical Composition of *Costus Afer* Extract and Its Alleviation 122 of Carbon Tetrachloride – induced Hepatic Oxidative Stress and Toxicity. *International Journal of Modern Botany*. 2012;2:120-126.
24. Hemali P, Anjali MT, Sumitra C. Antimicrobial activity of some medicinal plant extracts and its synergistic interaction with some antibiotics. *Journal of Pharmacy Research*. 2016;10 (5): 211-220
25. Baskar R, Rajeswari V, Kumar TS. In-vitro antioxidant studies in leaves of *Annona species* *Indian Journal of Experimental Biology*. 2007;45(5) 480-485.
PMID: 17569293.
26. Houcine B, Asma B, Faiza M, Abdelillah A. Free radical scavenging activity, kinetic behavior and phytochemical constituents of *Aristolochia clematiti* L. roots. *Arabian Journal of Chemistry*. 2017;10(1):S1402-S1408.

- Available:<http://dx.doi.org/10.1016/j.arabjc.2013.04.015>
27. Okawa M, Kinjo J, Nohara T, and Ono, M. DPPH (1, 1-diphenyl-2-picrylhydrazyl) radical scavenging activity of flavonoids obtained from some medicinal plants. *Biological and Pharmaceutical Bulletin*: 2001;24:1202-1205.
28. Zhiyong C, Yuanzong L. Reducing power: the measure of antioxidant activities of reductant compound. *Redox Report*. 2004;9:4:213-217.
Available:<https://doi.org/10.1179/135100004225005994>
29. Satish BN, Dilipkumar P. Free radicals, natural antioxidants, and their reaction mechanisms. *Royal Society of Chemistry*. 2015;5:27986-28006.
Available:<https://doi.org/10.1039/C4RA13315C>
30. Benzie IF, Strain JJ . The ferric reducing ability of plasma (FRAP) as a measure of antioxidant power: The FRAP assay. *Analytical Biochemistry*. 1996;239:70–76.
31. Kalu O, Igwe CO, Ujowundu SC, Chukwudoruo, Obasi UK. Assessment of hematological and serum electrolyte of albino rat administered with graded concentration of ethanol extract of *ficus capensis*. *Asian Journal of research in Botany*. 2020;4(3):28-36
32. Mensor LL, Fabio SM, Gilda GL, Alexandre SR, Teredza CD, Cintia SC, Suzana GL. Screening of Brazilian plant extracts for antioxidant activity by the use of DPPH free radical method. *Phytotherapy Research*. 2001;15:127-130.
Available: <https://doi.org/10.1002/ptr.687>
33. Omokhua GE. Medicinal and socio-cultural importance of *Costus Afer* (Ker Grawl) in Nigeria. *African Research Review*. 2011;5:282–7.
34. Katia A, Sara B, Simone C, Giuseppe R, Maria RC. Nitric oxide is the primary mediator of cytotoxicity induce GSH depletion in neuronal cells. *Journal of Cell Science*. 2011;124:1043-1054

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